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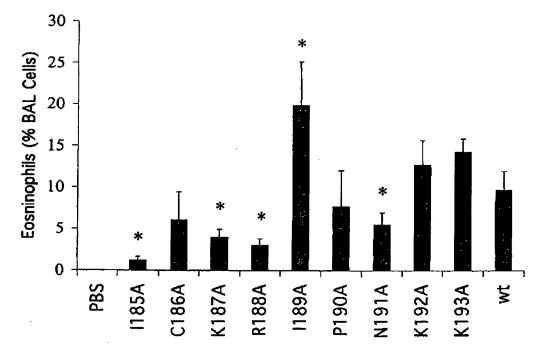
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[Continued on next page]

(54) Title: SUBUNIT VACCINE AGAINST RESPIRATORY SYNCYTIAL VIRUS INFECTION



(57) Abstract: The present invention relates generally to methods of treating or preventing RSV infections, and more specifically, to compositions, and the use thereof, comprising one or more RSV G protein immunogen or fragment thereof capable of eliciting protective immunity without eliciting an immunopathological response or eliciting a reduced immunopathological response.

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SUBUNIT VACCINE AGAINST RESPIRATORY SYNCYTIAL VIRUS INFECTION

CROSS-REFERENCE TO RELATED APPLICATIONS

This application claims the benefit of U.S. Provisional Patent

Application No. 60/567,586, filed May 3, 2004; and U.S. Provisional Patent

Application No. 60/487,804, filed July 15, 2003, in which these provisional applications are incorporated herein by reference in their entireties.

TECHNICAL FIELD

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The present invention relates generally to the prevention of infectious disease, and more specifically, to compositions, and the use thereof, comprising one or more respiratory syncytial virus G protein immunogens and fragments or variants thereof capable of eliciting protective immunity without eliciting an immunopathological response or with a reduced immunopathological response (e.g., reducing associated pulmonary pathology).

15 BACKGROUND

Respiratory syncytial virus (RSV) is the leading cause of lower respiratory tract infection (acute bronchiolitis and pneumonia) in early infancy (Glezen et al., Amer. J. Dis. Child. 140:543, 1986; Holberg et al., Am. J. Epidemiol. 133:1135, 1991; "Fields Virology", Fields, B. N. et al. Raven Press, N. Y. (1996), particularly, Chapter 44, pp 1313-1351 "Respiratory Syncytial Virus" by Collins, P., McIntosh, K., and Chanock, R. M.). Virtually all children are infected with RSV by the age of two years and 1-2% of all infected children require hospitalization (Holberg et al.; Parrott et al., Am. J. Epidemiol. 98:289, 1973). Outbreaks of RSV infection and lower respiratory tract deaths in infants and young children show a strong correlation (Anderson et al., J. Infect. Dis. 161:640, 1990), and mortality rates among hospitalized children range between 0.1-1% in the U.S. and Canada (Holberg et al.; Parrott et al.; Navas et al., J. Pediatr. 121:348, 1992; Law et al., Pediatr. Infect. Dis. J. 12:659, 1993; Ruuskanen and Ogra, Curr. Prob. Pediatr. 23:50, 1993). The consequences of RSV infection

during infancy range from bronchiolitis or pneumonia to an increased risk for childhood asthma.

Despite intense efforts spanning the past four decades, the search for a safe and effective vaccine against RSV remains elusive. Initial RSV vaccines, including formalin-inactivated and live attenuated virus (reviewed in Murphy et al., Virus Res. 32:13, 1994), proved to be disappointingly non-protective and actually led to more severe lung disease in vaccinated children who subsequently acquired natural RSV infection. Immunopathological responses, especially involving inflammatory cell infiltration, may likely underlie RSV-mediated damage to lung tissue. Children who received the formalin-inactivated RSV vaccine developed high levels of virus-specific antibodies, but the antibodies had low levels of neutralizing activity (Murphy et al., J. Clin. Microbiol. 24:197, 1986) and failed to protect against infection by RSV (Kim et al., Am. J. Epidemiol. 89:422, 1969; Kapikian et al., Am. J. Epidemiol. 89:405, 1969; Fulginiti et al., Am. J. Epidemiol. 89:435, 1969; Chin et al., Virol. 1:1, 1969).

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More recent efforts for development of an RSV vaccine have focused on subunit and recombinant methods. RSV has two major surface glycoproteins (designated F and G), which have been examined for use in potential vaccines. The F protein is involved in membrane fusion between the virus and target cell (Walsh and Hruska, J. Virol. 47:171, 1983), whereas the G protein is thought to mediate attachment of the virus to a cell receptor (Levine et al., J. Gen. Virol. 68:2521, 1987). Both RSV F and G proteins induce strong serum and mucosal immunity, which are important for protection against RSV infection (Glezen et al., 1986; Holberg et al.; Glezen et al., J. Pediatr. 98:708, 1981; Lamprecht et al., J. Infect. Dis. 134:211, 1976; Hemming et al., Clin. Microbiol. Rev. 8:22, 1995). Studies with mice have demonstrated that formalininactivated RSV and some G protein-encoding vaccinia recombinants prime for a harmful lung inflammatory response in which eosinophils are a prominent participant (Connors et al., J. Virol. 68:5321, 1994; Doherty, Trends Microbiol. 2:148, 1994; Waris et al., J. Virol. 70:2852, 1996; Graham et al., J. Immunol. 151:2032, 1993; Beasley et al., Thorax 43:679, 1988; Openshaw et al., Int. Immunol. 4:493, 1992).

Eosinophils and the eosinophil-attractant cytokine IL-5 are considered to be a feature of the so-called type 2 immune response, which has fostered the idea that

immunization with RSV antigen has the potential to trigger type 2 responses depending on factors, such as the nature of specific viral immunogens and their route of presentation (Openshaw et al., 1992; Kakuk et al., J. Infect. Dis. 167:553, 1993; Openshaw and O'Donnell, Thorax 49:101, 1994). Recent work indicates that a portion of the conserved region of the RSV G protein is involved in protective immunity against RSV and in the generation of inflammatory responses, including the induction of eosinophilia (Sparer et al., J. Expt'l. Med. 187:1921, 1998; Tebbey et al., J. Expt'l. Med. 188:1967, 1998; Srikiatchachorn et al., J. Virol. 73:6590, 1999; Varga et al., J. Immunol. 165:6487, 2000; Huang and Anderson, Vaccine 21:2500, 2003).

Hence, a need exists for identifying and developing compositions therapeutically effective against RSV infections, particularly those compositions that can function as a vaccine by eliciting protective immunity without any or with a reduced associated harmful pulmonary inflammation. Furthermore, there is a need for vaccine formulations that can be varied to protect against or treat for infection by different RSV immunogenic subtypes and subgroups. The present invention meets such needs, and further provides other related advantages.

BRIEF SUMMARY OF THE DISCLOSURE

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The present invention provides the discovery of therapeutic formulations of respiratory syncytial virus (RSV) immunogens, particularly G protein immunogens useful for eliciting a protective immune response without eliciting an, or with a reduced, immunopathological response.

In one aspect, the invention provides a method for treating or preventing an RSV infection, comprising administering to a subject in need thereof a composition comprising at least one respiratory syncytial virus G protein immunogen or fragment thereof comprising an amino acid sequence that is at least 80% identical to SEQ ID NO:2, wherein said G protein immunogen has an epitope that elicits a protective immune response without eliciting an, or with a reduced, immunopathological response, and a pharmaceutically acceptable carrier, diluent, or excipient, at a dose sufficient to elicit an immune response specific for one or more G protein immunogen or fragments and variants thereof. In a related embodiment, the G protein immunogen is an amino

acid sequence comprising or consisting of SEQ ID NO:2. In other embodiments, the invention provides a method for treating or preventing a respiratory syncytial virus infection wherein the G protein immunogen comprises an amino acid sequence selected from SEQ ID NOS:6, 8, 10, 12, 14,16, 18, 20, 22, 56, 58, 60, 62, 64, 66, 68 or 70. In still more embodiments, the invention provides a method wherein the composition further comprises at least one respiratory syncytial virus F protein immunogen or M protein immunogen, wherein the F protein and M protein immunogens have an epitope that elicits a protective immune response without eliciting an, or with a reduced, immunopathological response, or has at least two G protein immunogens.

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In another embodiment, any of the aforementioned G protein immunogens and fragments or variants thereof further comprise a hydrophobic portion or moiety (e.g., to act as an anchor or foot in a lipid environment such as a membrane or proteosome or liposome), particularly when formulated with a proteosome adjuvant delivery vehicle. In yet other embodiments, the hydrophobic mojety comprises an amino acid sequence or a lipid. In certain embodiments, the carrier is a liposome, and in other embodiments the liposome contains Deinococcus radiodurans lipids or α-galactosylphosphotidylglycerol alkylamine. In another embodiment, any of the aforementioned compositions further comprise an adjuvant, such as alum, Freund's adjuvant, or a proteosome-based formulation (e.g., a proteosome adjuvant delivery system). Preferably, the adjuvant is suitable for use in humans. In other embodiments, the G protein immunogen or fragment and variants thereof further comprise a second amino acid sequence to form a fusion protein, wherein the second amino acid sequence can be a tag, an enzyme or a combination thereof, such as a polyhistidine, thioredoxin, or both. In certain embodiments, such fusion proteins may further comprise a hydrophobic moiety. In yet other embodiments, any of the aforementioned methods are provided for use when the immunopathological response resulting from or associated with RSV infection is eosinophilia (such as pulmonary eosinophilia) or asthma. In still more embodiments, the invention provides any of the aforementioned methods for use when the infection is due to an RSV of subgroup A, subgroup B, or both subgroup A and subgroup B. In related embodiments, any of the disclosed compositions may be

administered in any of the aforementioned methods by a route selected from enteral, parenteral, transdermal, transmucosal, nasal or inhalation.

In another aspect, the invention provides a plurality of antibodies, Th cells, or both produced by a method according to any one of aforementioned methods.

In one embodiment, there is provided a method for treating or preventing an RSV infection, comprising administering to a subject in need thereof a composition comprising a pharmaceutically acceptable carrier or a proteosome adjuvant delivery vehicle, and a plurality of antibodies as just described.

In still another aspect, there is provided a composition comprising a respiratory syncytial virus G protein immunogen formulated with a proteosome adjuvant delivery vehicle, wherein said G protein immunogen comprises an amino acid sequence that is at least 80% identical to SEQ ID NO:2 or fragment thereof and wherein said G protein immunogen or fragment thereof has an epitope that elicits a protective immune response without eliciting an immunopathological response or with a reduced immunopathological response. In other embodiments, the composition includes any of the aforementioned G protein immunogens and fragments or variants thereof, fusion proteins, multivalent fusions, cocktail compositions or any combination thereof, and other additives, such as an adjuvant. In some embodiments, the adjuvant is alum, proteosome or protollin.

These and other aspects of the present invention will become evident upon reference to the following detailed description and attached drawings.

BRIEF DESCRIPTION OF THE DRAWINGS

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Figure 1 shows the induction of serum antibodies in mice immunized with wild type or mutant Trx-(polyHis)-G(128-229) proteins in alum and their ability to recognize wild type RSV G protein and the effect of G protein mutations on such induction. Extracts of RSV-infected HEp-2 cells were resolved by SDS-PAGE, transferred to membranes, and probed with pooled sera from each group of mice (mice were immunized twice at 14-day intervals with PBS/alum, or alum-adjuvanted wild type or mutant Trx-(polyHis)-G(128-229) proteins). Shown is an immunoblot of an

SDS-PAGE gel illustrating the specificity of mouse sera (1:100 dilution) for the RSV G protein.

Figures 2A and 2B show how G protein variants affect protective immunity (A) and eosinophilic infiltration in bronchoalveolar fluids (B) in immunized mice challenged with RSV. Mice were immunized twice subcutaneously at 14-day intervals with PBS/alum or wild type or mutant Trx-G128-229 proteins in alum, followed by RSV challenge. RSV titers in lung homogenates, as well as bronchoalveolar lavage eosinophils (as % of total cells) were determined four days after RSV challenge. Results are shown as means ± SD.

Figures 3A and 3B show the nucleic acid sequence (SEQ ID NO:1) and amino acid sequence (SEQ ID NO:2) of RSV group A, Long strain G protein. Shown in bold is an exemplary mutation of an amino acid (N191A, from codon AAC to GCC) to generate a G protein immunogen of the invention, from which fragments and variants thereof can be used as described herein.

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Figures 4A and 4B show polyacrylamide gel autoradiograms of ribonuclease protection assays (RPAs) of cytokine mRNA in lung tissue. The results illustrate relative levels of cytokine mRNA in lungs of mice assayed four days after RSV challenge, having been previously immunized twice subcutaneously at 14-day intervals with PBS/alum alone, alum-adjuvanted wild type Trx-G128-229 protein, or variant Trx-G128-229 proteins. Panels A and B show different regions of the polyacrylamide gel that exposed radiographic film for 3 days (A) or 1 hour (B).

Figure 5 shows the detection by ELISA of specific serum IgG antibodies from BALB/c mice immunized with wild type or mutant Trx-(polyHis)-G(128-229) fusion proteins alone, or adjuvanted with protollin or alum. Mice were immunized three times with a dose of 6 μ g or 2 μ g of Trx-(polyHis)-G(128-229) fusion proteins. Protollin alone or fusion proteins formulated with protollin were administered intranasally, and alum alone or fusion proteins formulated with alum were administered subcutaneously. Serum samples were obtained after the second immunization (day 35) and two weeks after the third immunization (day 62).

Figure 6 shows the detection by ELISA of specific bronchoalveolar lavage (BAL) IgA antibodies from BALB/c mice immunized with wild type or mutant

Trx-(polyHis)-G(128-229) fusion proteins alone, or adjuvanted with protollin or alum. Mice were immunized three times with a dose of 6 µg or 2 µg of Trx-(polyHis)-G(128-229) fusion proteins. Protollin alone or fusion proteins formulated with protollin were administered intranasally, and alum alone or fusion proteins formulated with alum were administered subcutaneously. BAL samples were collected on day 62 (two weeks after the third immunization).

DETAILED DESCRIPTION

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As set forth above, the present invention provides compositions and methods for using and making respiratory syncytial virus (RSV) G protein immunogen to treat or prevent respiratory syncytial virus infection. Although protection against RSV re-infection (i.e., challenge) could be obtained with previous vaccines consisting of various forms and immunization modes of the RSV G protein, this was often associated with an unwanted and harmful pulmonary inflammation characterized by pronounced eosinophilia. In addition, immunization of subjects prone to serious RSV disease (e.g., human subjects between the ages of 2 and 7 months of age) may be difficult due to possible immunosuppressive effects of maternally derived serum RSVneutralizing antibodies or because of the immunological immaturity of the subject. The instant invention, therefore, relates generally to the surprising discovery that certain RSV G protein fragments can be modified to induce or elicit protective immunity against RSV and not induce or have a reduced level of a concomitant immunopathological event that leads to, for example, pulmonary inflammation and aggravated disease upon subsequent infection with RSV. In particular, these G protein immunogens are useful for treating or preventing infections involving RSV. Discussed in more detail below are G protein immunogens or fragments and variants thereof suitable for use within the present invention, as well as representative compositions and therapeutic uses.

In the present description, any concentration range, percentage range, ratio range or integer range is to be understood to include the value of any integer within the recited range and, when appropriate, fractions thereof (such as one tenth and one hundredth of an integer), unless otherwise indicated. As used herein, "about" or

"comprising essentially of" mean ± 15%. The use of the alternative (e.g., "or") should be understood to mean either one, both or any combination thereof of the alternatives. In addition, it should be understood that the individual compounds, or groups of compounds, derived from the various combinations of the sequences, structures, and substituents described herein, are disclosed by the present application to the same extent as if each compound or group of compounds was set forth individually. Thus, selection of particular sequences, structures, or substituents is within the scope of the present invention.

RSV G PROTEIN IMMUNOGENS

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The present invention is directed generally to immunogenic RSV polypeptide immunogens of G protein or fragments and variants thereof, including fusions to other polypeptides (e.g., a tag, another protein, a hydrophobic amino acid sequence, or any combination thereof) or other modifications (e.g., addition of a lipid or glycosylation). The immunogenic G polypeptides may comprise any portion or fragment of a G protein that has an epitope capable of eliciting a protective immune response against RSV infection without eliciting an immunopathological response or with a reduced immunopathological response. Immunogenic polypeptides of the instant invention may be arranged or combined in a linear form, and each immunogen may or may not be reiterated, wherein the reiteration may occur once or multiple times. In addition, a plurality of different RSV immunogenic polypeptides (e.g., different G protein, F protein, or M protein variants and fragments or variants thereof) can be selected and mixed or combined into a cocktail composition or fused, conjugated or linked to provide a multivalent vaccine for use in eliciting a protective immune response without a harmful associated immune response.

As used herein, "G protein immunogen" or "RSV immunogen" refers to all full length polypeptides, full length variants, fragments and variants thereof, multivalent fusions, cocktail compositions, fusion proteins, or any combination thereof, capable of eliciting a protective immune response against RSV infection without eliciting an immunopathological response or with a reduced immunopathological response, as described herein.

The present invention further provides methods for producing synthetic or recombinant multivalent RSV polypeptide immunogens, including fusion proteins. For example, host cells containing G protein immunogen-encoding nucleic acid expression constructs may be cultured to produce recombinant G protein immunogens and fragments or variants thereof. Also contemplated are methods for treating or preventing RSV infections or eliciting an immune response using a G protein immunogens and fragments or variants thereof, or a combination of polypeptides (including fusion proteins).

As used herein, the phrase "immunopathological response" refers to a condition or disease resulting from an immune reaction, which may or may not have 10 detectable clinical symptoms. Exemplary immunopathological responses include hypersensitivity or asthma. Another exemplary immunopathological response can be an atypical induction of granulocytes in response to type 2 cytokines, such as is found in blood eosinophilia or pulmonary eosinophilia, which can be characteristic of an allergic state or a microbial infection (such as a parasitic infection or a respiratory syncytial virus infection).

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By way of background and not wishing to be bound by theory, RSV has a negative-sense, non-segmented, single-stranded RNA genome, which encodes at least 10 viral proteins (G, F, SH, M, M2, N, P, L, NS1, and NS2). RSV has two major surface glycoproteins (designated F and G), which have been examined for use in potential vaccines. The F protein is involved in membrane fusion between the virus and target cell (Walsh and Hruska, J. Virol. 47:171, 1983), whereas the G protein is thought to mediate attachment of the virus to a cell receptor (Levine et al., J. Gen. Virol. 68:2521, 1987). Both RSV F and G proteins induce strong serum and mucosal immunity, which are important for protection against RSV infection (Glezen et al., 1986; Holberg et al.; Glezen et al., J. Pediatr. 98:708, 1981; Lamprecht et al., J. Infect. Dis. 134:211, 1976; Hemming et al., Clin. Microbiol. Rev. 8:22, 1995). Studies with mice have demonstrated that formalin-inactivated RSV and some G protein-encoding vaccinia recombinants prime for a harmful lung inflammatory response in which eosinophils are a prominent participant (Connors et al., J. Virol. 68:5321, 1994; Doherty, Trends Microbiol. 2:148, 1994; Waris et al., J. Virol. 70:2852, 1996; Graham

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et al., J. Immunol. 151:2032, 1993; Beasley et al., Thorax 43:679, 1988; Openshaw et al., Int. Immunol. 4:493, 1992). A surprising result of the instant invention is the identification of G protein immunogens (e.g., variants and mutants of wild-type G protein; an exemplary wild-type G protein is set forth in SEQ 1D NO:4, which can be 5 encoded by a nucleic acid sequence as set forth in SEQ ID NO:3) that elicit a protective immune response without eliciting an, or with a reduced, immunopathological response. Thus, in certain embodiments of the instant invention, a respiratory syncytial virus G protein immunogen or fragment thereof that has an epitope that elicits a protective immune response without eliciting an, or with a reduced, immunopathological response is used to prepare compositions useful for treating or preventing RSV infections.

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In certain embodiments, the RSV G protein immunogens have at least 50% to 100% amino acid identity to an amino acid sequence of the full length G protein mutant as set forth in SEQ ID NO:2 (from RSV Group A, Long strain; SEQ ID NO:1 is the nucleic acid sequence that encodes amino acid sequence of SEQ ID NO:2), or fragments thereof as set forth in SEQ ID NOS:6, 8, 10, 12, 14,16, 18, 20, 22, 56, 58, 60, 62, 64, 66, 68 or 70; preferably 60%-99% identity, more preferably 70%-97% identity, and most preferably 80%-95% identity, wherein the G protein immunogen variants retain at least one epitope that elicits a protective immune response against RSV without eliciting an immunopathological response or with a reduced immunopathological response (e.g., eosinophilia). As used herein, "percent identity" or "% identity" is the percentage value returned by comparing the whole of the subject polypeptide, peptide, or variant thereof sequence to a test sequence using a computer implemented algorithm, typically with default parameters.

In one preferred embodiment, a G protein immunogen is a variant of wild-type G protein having a point mutation, wherein an amino acid at, for example, position 191 (Asn) is changed to an Ala (see SEQ ID NO:2 and Figure 3, also referred to as G protein N191A), and in a more preferred embodiment, a G protein immunogen variant is a fragment of full length G protein. For example such a G protein fragment may include from about amino acid 128 to about amino acid 229, wherein the fragment contains the N191A mutation (SEQ ID NO:6). In other embodiments, G protein immunogen variants span amino acids 128 to 229, wherein the variants include double

point mutants, such as P190A and N191A (SEQ ID NO:56), or R188A and N191A (SEQ ID NO:58). Other point mutants of use in the instant invention could include those at the Asn at positions 178 (SEQ ID NO:60) and 179 (SEQ ID NO:62), and at the Lys at positions 196 (SEQ ID NO:64), 197 (SEQ ID NO:66), 204 (SEQ ID NO:68), or 205 (SEQ ID NO:70).

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The representative G protein immunogen variants described herein include an Ala substitution, but the invention is not so limited and a person of skill in the art would know that other amino acids could be used for substitutions. Moreover, the variant immunogens of the instant invention could be made to include one or more of a variety of mutations, such as point mutations, frameshift mutations, missense mutations, additions, deletions, and the like, or the variants can be a result of modifications, such as by certain chemical substituents, including glycosylation, alkylation, etc. Each of the variants of the instant disclosure preferably is capable of eliciting a protective immune response against RSV without eliciting an immunopathological response or with a reduced immunopathological response (e.g., eosinophilia).

As described herein, preferred fragments of G protein, whether derived from RSV group A or group B, are immunogens that retain at least one epitope that elicits a protective immune response against RSV and elicits a reduced immunopathological response, or is incapable of eliciting an immunopathological response. In certain embodiments, the immunogen fragments or variants thereof (e.g., the N191A mutation) have mutations or variations from wild-type G protein in amino acid sequences that span from about amino acid 120 to about amino acid 300 of SEQ ID NO:2, preferably from about amino acid 125 to about amino acid 250, more preferably from about amino acid 150 to about amino acid 225, and most preferably from about amino acid 165 to about amino acid 195. In one embodiment, the G protein immunogen fragment includes amino acids 128 to 229 and mutations can be found in the range of about amino acids 178 to about 205 of G protein.

Sequence comparisons can be performed using any standard software program, such as BLAST, tBLAST, pBLAST, or MegAlign. Still others include those provided in the Lasergene bioinformatics computing suite, which is produced by

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DNASTAR® (Madison, Wisconsin). References for algorithms such as ALIGN or BLAST may be found in, for example, Altschul, J. Mol. Biol. 219:555-565, 1991; or Henikoff and Henikoff, Proc. Natl. Acad. Sci. USA 89:10915-10919, 1992. BLAST is available at the NCBI website (www.ncbi.nlm.nih.gov/BLAST). Other methods for comparing multiple nucleotide or amino acid sequences by determining optimal alignment are well known to those of skill in the art (see, e.g., Peruski and Peruski, The Internet and the New Biology: Tools for Genomic and Molecular Research (ASM Press, Inc. 1997); Wu et al. (eds.), "Information Superhighway and Computer Databases of Nucleic Acids and Proteins," in Methods in Gene Biotechnology, pages 123-151 (CRC Press, Inc. 1997); and Bishop (ed.), Guide to Human Genome Computing, 2nd Edition, Academic Press, Inc., 1998).

As used herein, "similarity" between two peptides or polypeptides is generally determined by comparing the amino acid sequence of one peptide or polypeptide to the amino acid sequence and conserved amino acid substitutes thereto of a second peptide or polypeptide. Fragments or portions of the G protein immunogens or variants thereof of the present description may be employed for producing the corresponding full-length G protein immunogens by peptide synthesis; therefore, the fragments may be employed as intermediates for producing the full-length G protein immunogens. Similarly, fragments or portions of the nucleic acids of the present invention may be used to synthesize full-length nucleic acids of the present disclosure.

As described herein, G protein immunogens and fragments or variants thereof of the instant disclosure have an epitope that elicits a protective immune response without eliciting an immunopathological response or with a reduced immunopathological response. The fragments and variants may be identified using *in vivo* and *in vitro* assays known in the art, such as animal immunization studies (e.g., using a mouse or rabbit model) and Western immunoblot analysis, respectively, and combinations thereof. Other examples include plaque reduction assays to assess whether G protein immunogens and fragments or variants thereof of the instant description are capable of eliciting an immune response, particularly a protective (neutralizing) immune response. Briefly, an animal is immunized with one or more G protein immunogens, or composition thereof, by subcutaneous administration, sera is

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collected from the immunized animals, and then the sera is tested for its ability to inhibit RSV infection of a cell culture monolayer (infection being measured as the number of plaques that form; i.e., "holes" in the monolayer arising from RSV causing cells to lyse) (see, e.g., Example 8). In addition, altered (reduced or enhanced) immunopathological responses can be indirectly identified by, for instance, examining cytokine expression patterns in animals challenged with RSV after immunization with G protein immunogens of the invention. For example, specific cytokine levels can be measured in tissues of interest using a ribonuclease protection assay (RPA) to deduce whether a type 1 or type 2 response is prevalent after immunization with a G protein 10 immunogen of the invention and subsequent challenge with RSV (see Example 9). These and other assays known in the art can be used to identify G protein immunogens and fragments or variants thereof that have an epitope that elicits a protective immune response without eliciting an immunopathological response or with a reduced immunopathological response, according to the instant description.

The RSV G protein polypeptides, fragments thereof, and fusion proteins thereof, as well as corresponding nucleic acids of the present invention, are preferably provided in an isolated form, and in certain preferred embodiments, are purified to homogeneity. As used herein, the term "isolated" means that the material is removed from its original or natural environment. For example, a naturally occurring nucleic acid molecule or polypeptide present in a living animal or cell is not isolated, but the same nucleic acid molecule or polypeptide is isolated when separated from some or all of the co-existing materials in the natural system. The nucleic acid molecules, for example, could be part of a vector and/or such nucleic acids or polypeptides could be part of a composition and still be isolated in that such vector or composition is not part of its natural environment.

The present invention also pertains to RSV G protein immunogens and fragments or variants thereof produced synthetically or recombinantly, and preferably recombinantly. The immunogenic polypeptide components of the immunogens may be synthesized by standard chemical methods, including synthesis by automated procedure. In general, immunogenic peptides are synthesized based on the standard solid-phase Fmoc protection strategy with HATU as the coupling agent. The

immunogenic peptide is cleaved from the solid-phase resin with trifluoroacetic acid containing appropriate scavengers, which also deprotects side chain functional groups. Crude immunogenic peptide is further purified using preparative reverse phase chromatography. Other purification methods, such as partition chromatography, gel filtration, gel electrophoresis, or ion-exchange chromatography may be used. Other synthesis techniques known in the art may be employed to produce similar immunogenic peptides, such as the tBoc protection strategy, use of different coupling reagents, and the like. In addition, any naturally occurring amino acid or derivative thereof may be used, including D- or L-amino acids and combinations thereof. In particularly preferred embodiments, a synthetic G protein immunogen of the invention will have an amino acid sequence that is at least 80% identical to SEQ ID NOS:2, 6, 8, 10, 12, 14,16, 18, 20, 22, 56, 58, 60, 62, 64, 66, 68 or 70.

As described herein, the G protein immunogens and fragments or variants thereof of certain embodiments may be recombinant, wherein a desired G protein immunogen is expressed from a polynucleotide that is operably linked to an expression control sequence (e.g., promoter) in a nucleic acid expression construct. In particularly preferred embodiments, a recombinant G protein immunogen will comprise an amino acid sequence that is at least 80% identical to SEQ ID NO:2. Some preferable recombinant G protein immunogens comprise an amino acid sequence of SEQ ID NO:2 or consist solely of an amino acid sequence as set forth in SEQ ID NO:2. More 20 preferably, a recombinant G protein immunogens and variants thereof comprise an amino acid sequence as set forth in SEQ ID NOS:6, 8, 10, 12, 14,16, 18, 20, 22, 56, 58, 60, 62, 64, 66, 68 or 70, and more preferably comprise an amino acid sequence as set forth in SEQ ID NO:6, SEQ ID NO:56 or SEQ ID NO:58. In preferred embodiments, recombinant G protein immunogens and fragments or variants thereof have an epitope 25 that elicits a protective immune response without eliciting an, or with a reduced, immunopathological response.

"Nucleic acid" or "nucleic acid molecule" refers to any of deoxyribonucleic acid (DNA), ribonucleic acid (RNA), oligonucleotides, fragments generated by the polymerase chain reaction (PCR), and fragments generated by any of ligation, scission, endonuclease action, and exonuclease action. Nucleic acids may be

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composed of monomers that are naturally occurring nucleotides (such as deoxyribonucleotides and ribonucleotides), analogs of naturally occurring nucleotides (e.g., \alpha-enantiomeric forms of naturally-occurring nucleotides), or a combination of both. Modified nucleotides can have modifications in sugar moieties and/or in pyrimidine or purine base moieties. Sugar modifications include, for example, replacement of one or more hydroxyl groups with halogens, alkyl groups, amines, and azido groups, or sugars can be functionalized as ethers or esters. Moreover, the entire sugar moiety may be replaced with sterically and electronically similar structures, such as aza-sugars and carbocyclic sugar analogs. Examples of modifications in a base 10 moiety include alkylated purines and pyrimidines, acylated purines or pyrimidines, or other well-known heterocyclic substitutes. Nucleic acid monomers can be linked by phosphodiester bonds or analogs of such linkages. Analogs of phosphodiester linkages include phosphorothioate, phosphorodithioate, phosphoroselenoate, phosphorodiselenoate, phosphoroanilothioate, phosphoranilidate, phosphoramidate, and the like. The term "nucleic acid" also includes so-called "peptide nucleic acids," which comprise naturally occurring or modified nucleic acid bases attached to a polyamide backbone. Nucleic acids can be either single stranded or double stranded.

Further, an "isolated nucleic acid molecule" refers to a polynucleotide molecule in the form of a separate fragment, or as a component of a larger nucleic acid construct, which has been separated from its source cell (including the chromosome it normally resides in) at least once in a substantially pure form. For example, a DNA molecule that encodes an RSV polypeptide, peptide, or variant thereof, which has been separated from an RSV particle or from a host cell infected with or harboring RSV, is an isolated DNA molecule. Another example of an isolated nucleic acid molecule is a chemically synthesized nucleic acid molecule. Nucleic acid molecules may be comprised of a wide variety of nucleotides, including DNA, cDNA, RNA, nucleotide analogues or some combination thereof. In one embodiment, an isolated nucleic acid molecule comprises a sequence encoding a G protein immunogen or fragment thereof comprising an amino acid sequence that is at least 80% identical to SEQ ID NO:2, wherein said G protein immunogen has an epitope that elicits a protective immune response without eliciting an, or with a reduced, immunopathological response. In

another embodiment, an isolated nucleic acid molecule comprises a sequence encoding a G protein immunogen that has an amino acid sequence comprising or consisting of SEQ ID NO:2. In other embodiments, an isolated nucleic acid molecule comprises a sequence encoding a G protein immunogen fragment that comprises an amino acid sequence as set forth in NOS:6, 8, 10, 12, 14,16, 18, 20, 22, 56, 58, 60, 62, 64, 66, 68 or 70, and more preferably comprises an amino acid sequence as set forth in SEQ ID NO:6, SEQ ID NO:56, or SEQ ID NO:58.

In certain aspects, the invention relates to nucleic acid vectors and constructs that include nucleic acid sequences of the present invention, and in particular to "nucleic acid expression constructs" that include any polynucleotide encoding an RSV polypeptide and fragments or variants thereof as provided above. In another aspect, the instant disclosure pertains to host cells that are genetically engineered with vectors or constructs of the invention, and to the production and use in methods for treating or preventing an RSV infection or eliciting an immune response. The RSV polypeptides and fragments or variants thereof may be expressed in mammalian cells, yeast, bacteria or other cells under the control of appropriate expression control sequences. Cell-free translation systems may also be employed to produce such proteins using RNAs derived from the nucleic acid expression constructs of the present invention. Appropriate cloning and expression vectors for use with prokaryotic and eukaryotic hosts are described, for example, by Sambrook et al., Molecular Cloning: A Laboratory Manual, Second Edition, Cold Spring Harbor, NY, (1989), and may include plasmids, cosmids, shuttle vectors, viral vectors and vectors comprising a chromosomal origin of replication as disclosed therein.

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In one embodiment, a nucleic acid expression construct comprises an expression control sequence operably linked to a polynucleotide encoding a G protein immunogen or fragment thereof comprising an amino acid sequence that is at least 80% identical to SEQ ID NO:2, wherein said G protein immunogen has an epitope that elicits a protective immune response without eliciting an immunopathological response or with a reduced immunopathological response. In certain embodiments, a nucleic 30 acid expression construct comprises an expression control sequence operably linked to a polynucleotide encoding a G protein immunogen that has an amino acid sequence

comprising or consisting of SEQ ID NO:2. In other embodiments, a nucleic acid expression construct comprises an expression control sequence operably linked to a polynucleotide encoding a G protein immunogen or fragment thereof comprising an amino acid sequence as set forth in NOS:6, 8, 10, 12, 14,16, 18, 20, 22, 56, 58, 60, 62, 64, 66, 68 or 70, and more preferably comprises an amino acid sequence as set forth in SEQ ID NO:6, SEQ ID NO:56, or SEQ ID NO:58.

In other embodiments, the nucleic acid expression constructs described herein have an inducible promoter, which may be lac, tac, trc, ara, trp, \(\lambda \) phage, T7 phage, and T5 phage promoter, and more preferably is a T5 phage promoter/lac operator expression control sequence (plasmid pT5) as described in U.S. Patent Application Publication No. 2003/0143685. The "expression control sequence" refers to any sequence sufficient to allow expression of a protein of interest in a host cell, including one or more promoter sequences, enhancer sequences, operator sequences (e.g., lacO), and the like. In certain embodiments, the RSV polypeptide-encoding nucleic acid is in 15 a plasmid, preferably in plasmid pT5, and the host cell is a bacterium, preferably Escherichia coli.

Injection of mammals with gene delivery vehicles (e.g., naked DNA) encoding antigens of various pathogens has been shown to result in protective immune responses (Ulmer et al., Science 259:1745-9, 1993; Bourne et al., J Infect. Dis. 173:800-7, 1996; Hoffman et al., Vaccine 12:1529-33, 1994). Since the original description of in vivo expression of foreign proteins from naked DNA injected into muscle tissue (Wolff et al., Science 247:1465-8, 1990), there have been several advances in the design and delivery of DNA for purposes of vaccination.

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The RSV vaccines described herein are ideally suited for delivery via naked DNA because antibodies ultimately establish protective immunity. For example, 25 within one embodiment, polynucleotide sequences that encode a G protein immunogen or fragment thereof are ligated into plasmids that are specifically engineered for mammalian cell expression (see, e.g., Hartikka et al., Hum Gene Ther 7:1205-17, 1996, which contains the promoter/enhancer element from cytomegalovirus early gene, the signal peptide from human tissue plasminogen activator and a terminator element from the bovine growth hormone gene). The RSV polypeptides can be cloned into a plasmid

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that is used to transfect human cell lines to assure recombinant protein expression. The plasmid may be propagated in bacteria, such as E. coli, and purified in quantities sufficient for immunization studies by cesium chloride gradient centrifugation. Animals, such as mice, can be immunized with, for example, 50 µg of an isolated recombinant plasmid in 50 ul saline intramuscularly (i.m.). Booster injections of the same dose may be further given at three and six week intervals after the initial injection.

A wide variety of other gene delivery vehicles can likewise be utilized within the context of the present invention, including viruses, retrotransposons and cosmids. Representative examples include adenoviral vectors (e.g., WO 94/26914, WO 93/9191; Yei et al., Gene Therapy 1:192-200, 1994; Kolls et al., PNAS 91(1):215-219, 1994; Kass-Eisler et al., PNAS 90(24):11498-502, 1993; Guzman et al., Circulation 88(6):2838-48, 1993; Guzman et al., Cir. Res. 73(6):1202-1207, 1993; Zabner et al., Cell 75(2):207-216, 1993; Li et al., Hum Gene Ther. 4(4):403-409, 1993; Caillaud et al., Eur. J. Neurosci. 5(10):1287-1291, 1993), adeno-associated type 1 ("AAV-1") or adeno-associated type 2 ("AAV-2") vectors (see WO 95/13365; Flotte et al., PNAS 15 90(22):10613-10617, 1993), hepatitis delta vectors, live, attenuated delta viruses, vaccinia vectors and herpes viral vectors (e.g., U.S. Patent No. 5,288,641), as well as vectors which are disclosed within U.S. Patent No. 5,166,320. Other representative vectors include retroviral vectors (e.g., EP 0 415 731; WO 90/07936; WO 91/02805; WO 94/03622; WO 93/25698; WO 93/25234; U.S. Patent No. 5,219,740; WO 93/11230; WO 93/10218). Methods of using such vectors in gene therapy are well known in the art (see, e.g., Larrick, J.W. and Burck, K.L., Gene Therapy: Application of Molecular Biology, Elsevier Science Publishing Co., Inc., New York, New York, 1991; and Kreigler, M., Gene Transfer and Expression: A Laboratory Manual,

W.H. Freeman and Company, New York, 1990). 25

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Gene-delivery vehicles may be introduced into a host cell utilizing a vehicle, or by various physical methods. Representative examples of such methods include transformation using calcium phosphate precipitation (Dubensky et al., PNAS 81:7529-7533, 1984), direct microinjection of such nucleic acid molecules into intact target cells (Acsadi et al., Nature 352:815-818, 1991), and electroporation whereby cells suspended in a conducting solution are subjected to an intense electric field in

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order to transiently polarize the membrane, allowing entry of the nucleic acid molecules. Other procedures include the use of nucleic acid molecules linked to an inactive adenovirus (Cotton et al., PNAS 89:6094, 1990), lipofection (Felgner et al., Proc. Natl. Acad. Sci. USA 84:7413-7417, 1989), microprojectile bombardment

(Williams et al., PNAS 88:2726-2730, 1991), polycation compounds (such as polylysine), receptor specific ligands, liposomes entrapping the nucleic acid molecules, spheroplast fusion whereby E. coli containing the nucleic acid molecules are stripped of their outer cell walls and fused to animal cells using polyethylene glycol, viral transduction, (Cline et al., Pharmac. Ther. 29:69, 1985; and Friedmann et al., Science 244:1275, 1989), and DNA ligand (Wu et al, J. of Biol. Chem. 264:16985-16987, 1989), as well as psoralen inactivated viruses such as Sendai or Adenovirus.

Serum from a subject immunized with gene delivery vehicles containing RSV polypeptide immunogens and fragments or variants thereof, and fusions thereof can be assayed for total antibody titer by ELISA using native RSV polypeptides as the antigen. Serum protective antibodies may be assayed as described herein or as known in the art. Protective efficacy of DNA RSV polypeptide vaccines can be determined by, for example, direct animal protection assays (*i.e.*, challenge infection studies) using an RSV serotype that is represented in the pharmaceutical composition or vaccine (*i.e.*, challenge infection studies).

As will be appreciated by those of ordinary skill in the art, an RSV polypeptide-encoding nucleic acid may be a variant of the natural sequence due to, for example, the degeneracy of the genetic code (including homologs or strain variants or other variants). Briefly, such "variants" may result from natural polymorphisms or may be synthesized by recombinant methodology (e.g., to obtain codon optimization for expression in a particular host) or chemical synthesis, and may differ from wild-type polypeptides by one or more amino acid substitutions, insertions, deletions, and the like. Variants encompassing conservative amino acid substitutions include, for example, substitutions of one aliphatic amino acid for another, such as Ile, Val, Leu, or Ala or substitutions of one polar residue for another, such as between Lys and Arg, Glu and Asp, or Gln and Asn. Such substitutions are well known in the art to provide variants having similar physical properties, structural properties, and functional activities, such

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as for example, the ability to elicit and cross-react with similar antibodies (e.g., antibodies that specifically bind to wild-type G protein). Other variants include nucleic acids sequences that encode G protein immunogen fragments having at least 50% to 100% amino acid identity to SEQ ID NOS:2, 6, 8, 10, 12, 14,16, 18, 20, 22, 56, 58, 60, 62, 64, 66, 68 or 70. Preferred embodiments are those variants having greater than 90% or 95% identity with the amino acid sequence of SEQ ID NOS:2, 6, 8, 10, 12, 14,16, 18, 20, 22, 56, 58, 60, 62, 64, 66, 68 or 70.

In certain embodiments, the present invention includes any of the aforementioned degenerate nucleic acid molecules that encode G protein immunogens and fragments or variants thereof comprising an amino acid sequence as set forth in SEQ ID NOS:2, 6, 8, 10, 12, 14,16, 18, 20, 22, 56, 58, 60, 62, 64, 66, 68 or 70, and that retain functional activity (such as having an epitope that elicits a protective immune response without eliciting an immunopathological response or with a reduced immunopathological response). In another aspect, contemplated are nucleic acid molecules that encode G protein immunogens and fragments or variants thereof having conservative amino acid substitutions or deletions or substitutions, such that the RSV polypeptide variant(s) retain (from wild-type) or have at least one epitope capable of eliciting antibodies specific for one or more RSV strains.

In certain aspects, a nucleic acid sequence may be modified to encode an 20 RSV immunogen or functional variant thereof wherein specific codons of the nucleic acid sequence have been changed to codons that are favored by a particular host and can result in enhanced levels of expression (see, e.g., Haas et al., Curr. Biol. 6:315, 1996; Yang et al., Nucleic Acids Res. 24:4592, 1996). For example, certain codons of the immunogenic peptides can be optimized, without changing the primary sequence of the peptides, for improved expression in Escherichia coli. By way of illustration and not limitation, arginine (Arg) codons of AGG/AGA can be changed to the Arg codons of CGT/CGC. Similarly, AGG/AGA Arg codons can be optimized to CGT/CGC codons. As is known in the art, codons may be optimized for a host in which the G protein immunogens and fragments or variants thereof are to be expressed, including bacteria, fungi, insect cells, plant cells, and mammalian cells. Additionally, codons encoding different amino acids may be changed as well, wherein one or more codons

encoding different amino acids may be altered simultaneously as would best suit a particular host (e.g., codons for arginine, glycine, leucine, and serine may all be optimized or any combination thereof). Exemplary nucleic acid sequences with codons optimized for expression in bacteria include sequences as set forth in SEQ ID NOS:23, 25, 27, 29, 31 and 33. These nucleic acid sequences encode G protein immunogen fragment fusion proteins (i.e., fused to thioredoxin or a hexahistidine tag) as set forth in SEQ ID NOS:24, 26, 28, 30, 32 and 34, respectively. Alternatively, codon optimization may result in one or more changes in the primary amino acid sequence, such as a conservative amino acid substitution, addition, deletion, and combinations thereof.

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While particular embodiments of isolated nucleic acids encoding RSV immunogens are depicted in SEQ ID NOS:1, 3, 5, 7, 9, 11, 13,15, 17, 19, 21, 55, 57, 59, 61, 63, 65 67 or 69, within the context of the present disclosure, reference to one or more isolated nucleic acids includes variants of these sequences that are substantially similar in that they encode native or non-native RSV polypeptides with similar structure and similar functional ability to elicit specific antibodies to at least one G protein epitope contained in the RSV polypeptides of SEQ ID NOS:2, 4, 6, 8, 10, 12, 14,16, 18, 20, 22, 56, 58, 60, 62, 64, 66, 68 or 70. As used herein, the nucleotide sequence is deemed to be "substantially similar" when: (a) the nucleotide sequence is derived from the coding region of an RSV G protein gene (including, for example, portions of the sequence or homologous variations of the sequences discussed herein) and contains a G protein epitope with substantially the same ability to elicit a protective immune response without eliciting an, or with a reduced, immunopathological response; (b) the nucleotide sequence is capable of hybridization to the nucleotide sequences of the present invention under moderate or high stringency; (c) the nucleotide sequences are degenerate (i.e., sequences which code for the same amino acids using a different codon sequences) as a result of the genetic code to the nucleotide sequences defined in (a) or (b); or (d) is a complement of any of the sequences described in (a), (b) or (c).

As used herein, two nucleotide sequences are said to "hybridize" under conditions of a specified stringency when stable hybrids are formed between substantially complementary nucleic acid sequences. Stringency of hybridization refers to a description of the environment under which hybrids are annealed and washed (i.e.,

conditions under which annealed hybrids remain hybridized or annealed), which typically includes varying ionic strength and temperature. Other factors that might affect hybridization include the probe size and the length of time the hybrids are allowed to form. For example, "high," "medium" and "low" stringency encompass the following conditions or equivalent conditions thereto: high stringency is 0.1 x SSPE or SSC, 0.1% SDS, 65°C; medium stringency is 0.2 x SSPE or SSC, 0.1% SDS, 50°C; and low stringency is 1.0 x SSPE or SSC, 0.1% SDS, 50°C. As used herein, the term "high stringency conditions" means that one or more sequences will remain hybridized only if there is at least 95%, and preferably at least 97%, identity between the sequences. In preferred embodiments, the nucleic acid sequences that remain hybridized to a G protein immunogen-encoding nucleic acid molecule encode polypeptides that retain at least one epitope of a G protein immunogen of any one of SEQ ID NOS:2, 6, 8, 10, 12, 14,16, 18, 20, 22, 56, 58, 60, 62, 64, 66, 68 or 70, and have an epitope with substantially the same ability to elicit a protective immune response without eliciting an immunopathological response or with a reduced immunopathological response.

Methods for producing the RSV polypeptides of the subject invention are also provided wherein any of the nucleic acid molecules and host cells described herein may be used. In a preferred embodiment, a method of producing a G protein immunogen and fragments or variants thereof (having at least one epitope that elicits a protective immune response without eliciting an, or with a reduced, immunopathological response) comprises culturing a host cell containing a nucleic acid expression vector comprising at least one expression control sequence operably linked to a nucleic acid molecule encoding an RSV polypeptide, such as an RSV G protein immunogen and fragment or variant thereof as set forth in any one of SEQ ID NOS:2, 6, 8, 10, 12, 14,16, 18, 20, 22, 56, 58, 60, 62, 64, 66, 68 or 70, under conditions and for 25 a time sufficient for expression of the polypeptide. In one embodiment, an RSV G protein immunogen and fragment or variant thereof is produced by this method, and more preferably the RSV polypeptides produced comprise an amino acid sequence as set forth in SEQ ID NOS: 2, 6, 8, 10, 12, 14,16, 18, 20, 22, 56, 58, 60, 62, 64, 66, 68 or 70, and more preferably the RSV polypeptides produced comprise an amino acid 30 sequence as set forth in SEQ ID NO:6, 56, or 58.

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In certain embodiments, multivalent vaccines are contemplated. For example, such multivalent compositions may comprise a combination of two or more different G protein immunogens, or a combination of one or more G protein immunogens with one or more other RSV immunogens (such as an F protein or an M protein immunogen). The combination of antigens may be formulated as a cocktail (i.e., a mixture of a plurality of different immunogens), or the combination may be a plurality of different immunogens conjugated, linked or fused together (chemically or recombinantly). In addition, the fused immunogens may have one or more immunogens reiterated at least once within the multivalent fusion protein, which reiteration may occur at the amino-terminal end, the carboxy-terminal end, an internal position of a selected multivalent immunogen polypeptide, at multiple positions, or any combination thereof. For example, such multivalent hybrid RSV immunogens may comprise one or more peptide fragments of the G protein and one or more peptides fragments of an F protein or M protein of RSV, and any combination thereof. In certain embodiments, such multivalent hybrid RSV multivalent hybrid RSV immunogen vaccine compositions may combine immunogenic epitopes from different RSV antigenic groups, for example, immunogens from subgroup A viruses (e.g., Long and A2) or subgroup B viruses (e.g., CH-18537 and 8/60), or immunogens from both subgroup A and B viruses (or any other RSV subgroups that are found to, for example, infect humans).

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In some embodiments, the RSV immunogens may be linked by, for example, at least two amino acids encoded by a nucleic acid sequence that is a restriction enzyme recognition site, wherein the restriction sites may be any one or more of BamHI, ClaI, EcoRI, HindIII, KpnI, NcoI, NheI, PmII, PstI, SalI, XhoI, and the like. Additional amino acid linkers may also be added synthetically, as is known in the art and described herein. Preferably, the additional amino acids do not create any identity in sequence encompassing a five amino acid stretch of a human protein so as to minimize the possibility of eliciting human tissue cross-reactive antibodies. In addition, the hybrid polypeptides of the subject invention may further comprise at least one additional carboxy-terminal amino acid, wherein the additional amino acid is a D- or an L-amino acid. Any of the twenty naturally occurring amino acids or derivatives thereof

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may be added, such as cysteine, histidine, leucine, and glutamic acid. For example, the addition of cysteine may be useful to attach (e.g., enzymatically or by chemical cross-linking) other constituents, such as a lipid, a carrier protein, a tag, an enzyme, and the like.

As described herein, the invention also provides RSV immunogen fusion proteins comprising a G protein immunogen or fragment thereof fused to an additional functional or non-functional polypeptide sequence that permits, for example, detection, isolation, and purification of the hybrid polypeptide fusion proteins. For instance, an additional functional polypeptide sequence may be a tag sequence, which includes fusion proteins that may in certain embodiments be detected, isolated or purified by protein-protein affinity (e.g., receptor-ligand), metal affinity or charge affinity methods. In certain other embodiments the hybrid polypeptide fusion proteins may be detected by specific protease cleavage of a fusion protein having a sequence that comprises a protease recognition sequence, such that the hybrid polypeptides may be separable from the additional polypeptide sequence. In addition, the hybrid polypeptides may be made synthetically including additional amino acids, a carrier protein, a hydrophobic portion or moiety (e.g., a lipid), or a tag sequence, which may be located at the amino-terminal end, carboxy-terminal end, or at a site internal (non-terminal) of the fusion protein. In particularly preferred embodiments, for example, recombinant RSV immunogens are fused in-frame to a tag, which tag may be any one of alkaline phosphatase, thioredoxin (Trx), β-galactosidase, hexahistidine (6xHis), FLAG[®] epitope tag (DYKDDDDK, SEQ ID NO:71), GST or the like, and any combination thereof.

Preferred embodiments include hybrid polypeptide fusion proteins that facilitate affinity detection and isolation of the hybrid polypeptides, and may include, for example, poly-His or the defined antigenic peptide epitopes described in U.S. Patent No. 5,011,912 and in Hopp et al., (1988 *Bio/Technology* 6:1204), or the XPRESSTM epitope tag (DLYDDDDK, SEQ ID NO:72; Invitrogen, Carlsbad, CA), or thioredoxin. The affinity sequence may be a hexa-histidine tag as supplied by a vector. For example, a pBAD/His (Invitrogen), a pET vector (Invitrogen) or a pQE vector (Qiagen, Valencia, CA) can provide a polyhistidine tag for purification of the mature protein fusion from a particular host, such as a bacterium, using a nickel affinity column.

Alternatively, the affinity sequence may be added either synthetically or engineered into the primers used to recombinantly generate the nucleic acid sequence (e.g., using the polymerase chain reaction) encoding an immunogenic peptide of RSV. Optionally, any of the aforementioned G protein immunogens and fragments or variants thereof, and fusion proteins thereof, may also have a hydrophobic portion (anchor or foot) that is conjugated, linked or fused (chemically or recombinantly) to the amino-terminal end or carboxy-terminal end. Representative hydrophobic moieties include an amino acid sequence of at least five amino acids, such as MFLLAVFYGG (SEQ ID NO:35) or GGYFVALLF (SEQ ID NO:36), or a lipid.

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In certain embodiments, RSV immunogens are fused to a thioredoxin or a polyhistidine tag, which are encoded by a recombinant nucleic acid sequence encoding such a fusion protein. In preferred embodiments, RSV G protein immunogen fragments are fused to a thioredoxin and a polyhistidine tag, which are encoded by a nucleic acid sequence as set forth in SEQ ID NOS: 23 or 25. Exemplary amino acid sequences of RSV G protein immunogen fragments fused to a thioredoxin and a polyhistidine tag are set forth in SEQ ID NOS:24 and 26. In related embodiments, provided are nucleic acid sequences that encode an RSV G protein immunogen fusion protein further comprising a nucleic acid sequence that encodes a hydrophobic moiety or foot linked or fused to the G immunogen fusion protein, as found in the sequences set forth in SEQ ID NOS:27, 29, 31 or 33. Exemplary amino acid sequences of RSV G protein immunogen fragments fused to a thioredoxin and a polyhistidine tag, and further comprising a hydrophobic portion or foot are set forth in SEQ ID NOS:28, 30, 32 and 34. In preferred embodiments, the hydrophobic moiety is an amino acid sequence of MFLLAVFYGG (SEQ ID NO:35) fused to the amino-terminal end of the fusion protein or GGYFVALLF (SEQ ID NO:36) fused to the carboxy-terminal end of the fusion protein.

A fusion protein may comprise a hydrophobic moiety fused to the amino-terminal end or carboxy-terminal end of a G protein immunogen or fragment thereof. Alternatively, fusion protein may comprise a hydrophobic portion fused to a linker (e.g., one or more amino acids, preferably two or four) which in turn is fused to the amino-terminal end or carboxy-terminal end of a G protein immunogen or fragment

thereof. In still other embodiments, a fusion protein may comprise a hydrophobic moiety fused to one or more amino acid sequences (e.g., a tag, such as a thioredoxin or a polyhistidine) which in turn is fused to the amino-terminal end of a G protein immunogen or fragment thereof, or a fusion protein may comprise one or more amino acid sequences (e.g., a tag, such as a thioredoxin or a polyhistidine) fused to the amino-terminal end of a G protein immunogen or fragment thereof which in turn is fused to a hydrophobic portion. As will be appreciated by those of skill in the art, a fusion protein of the instant disclosure may be constructed to contain one or more G protein immunogens or fragments and variants thereof, one or more linkers, one or more additional amino acid tag sequences, one or more hydrophobic portions, or any combination thereof.

THERAPEUTIC FORMULATIONS AND METHODS OF USE

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This description also relates to pharmaceutical compositions that contain one or more RSV immunogens, which may be used to elicit an immune response without the concomitant immunopathological response or at least a reduced immunopathological response. This description further relates to methods for treating and preventing RSV infections by administering to a subject a G protein immunogen or fragment and variants thereof, fusion protein, multivalent immunogen, or a mixture of such immunogens at a dose sufficient to elicit antibodies specific for RSV, as described herein. G protein immunogens or fragments and variants thereof, or a cocktail of such immunogens are preferably part of a pharmaceutically acceptable composition when used in the methods of the present invention.

By way of background, natural or experimental infection of an animal or human subject does not appear to elicit a CD8⁺ CTL immune response recognizing G protein, while in contrast the F protein does elicit a CD8⁺ CTL immune response. Accordingly, a G plus F composite RSV antigen vaccine of the instant description is expected to elicit both a CD4⁺ and a CD8⁺ protective immune response, without eliciting an, or with a reduced, immunopathological proliferative lymphocyte response, such a response being harmful or otherwise unwanted. Moreover, the use of proteosome technology-based components combined with RSV antigen(s) may

influence a shift in the immune response raised to an RSV antigen from a predominantly type 2 response towards a preferential type 1 response (as determined by cytokine profiles known by those of ordinary skill in the art), and thereby eliminating or reducing, in a statistically significant manner, an undesired eosinophilic response, an undesired IgE response or both, following immunization with a vaccine or pharmaceutical composition of the instant description. For example, by combining one or more MHC class I CD8⁺ immunogenic epitopes of the RSV F protein with one or more CD4⁺ MHC class II immunogenic epitopes contained in RSV G protein.

10 lymphocyte activation at the time of first exposure to an RSV antigen influences the pattern of immune responses to subsequent exposures. Therefore, along with protection from respiratory disease and eosinophilia, immunogenic compositions of the instant application may prove to be useful in protecting against childhood asthma associated with an RSV infection. In one preferred embodiment, for example, a vaccine of the instant invention is capable of eliciting an immune response that protects from or otherwise moderates the pathological consequences of an RSV infection, while at the same time ablating or otherwise diminishing a subsequent IgE antibody response to common allergens.

In certain embodiments, the invention provides a composition

comprising a respiratory syncytial virus G protein immunogen formulated with a proteosome or a liposome, wherein said G protein immunogen comprises an amino acid sequence that is at least 80% identical to SEQ ID NO:2 or fragment thereof and wherein said G protein immunogen or fragment thereof has an epitope that elicits a protective immune response without eliciting an, or with a reduced, immunopathological response.

One embodiment is a G protein immunogen comprising an amino acid sequence as set forth in SEQ ID NO:2 or consisting of SEQ ID NO:2. In other preferred embodiments are G protein immunogens that comprise an amino acid sequence selected from SEQ ID NO:6, 8, 10, 12, 14,16, 18, 20, 22, 56, 58, 60, 62, 64, 66, 68 or 70, more preferably SEQ ID NO:6, SEQ ID NO:56 or SEQ ID NO:58. Preferably, liposomes formulated to contain one or more RSV immunogens further comprise *Deinococcus radiodurans* lipids or α-galactosylphosphotidylglycerolalkylamine. The addition of such lipids in a

liposome can enhance the efficacy of an RSV vaccine composition by increasing protective immunity and suppressing harmful eosinophilia (see, e.g., Huang and Anderson, *Vaccine 20*:1586, 2002).

Respiratory syncytial virus immunogens of the present invention may further include a covalently attached hydrophobic moiety. A hydrophobic moiety may be, for example, an amino acid sequence or a lipid, as disclosed in U.S. Patent No. 5,726,292. In certain embodiments, the hydrophobic moiety is an amino acid sequence of MFLLAVFYGG (SEQ ID NO:35) fused to the amino-terminal end of an immunogen or GGYFVALLF (SEQ ID NO:36) fused to the carboxy-terminal end of an immunogen. Naturally occurring RSV G protein contains a hydrophobic transmembrane amino acid sequence, which may function as a hydrophobic moiety according to the instant invention. In one embodiment, an RSV composition (e.g., a vaccine composition) of the instant application comprises an RSV G protein immunogen or fragment thereof as described herein formulated with a proteosome or protollin. When formulated with a proteosome or protollin, the G protein immunogens preferably further comprise a hydrophobic moiety, which may be composed of a hydrophobic amino acid sequence or a lipid (as used herein, lipid refers to a solubility characteristic and, therefore, includes alkyls, arylalkls, aryls, fatty acids, glycerides and glyceryl ethers, phospholipids, sphingolipids, long chain alcohols, steroids, vitamins, and the like). In certain embodiments, the G protein immunogens, with or without a 20 hydrophobic moiety, may further contain a second amino acid sequence to form a fusion, wherein the second amino acid sequence is a tag, carrier, enzyme or a combination thereof, as described herein. One preferred RSV vaccine of the instant invention can comprise a non-infectious RSV polypeptide or fragment thereof that is 25 highly immunogenic and capable of immunoneutralizing virus growth. In preferred embodiments of the instant invention, such an RSV subunit vaccine has reduced or no unwanted immunopathological side effects (e.g., eosinophilia or asthma) in a vaccinated subject, such as a human or animal.

The pharmaceutical composition will preferably include at least one of a

30 pharmaceutically acceptable vehicle, carrier, diluent, or excipient, in addition to one or
more RSV immunogen or fusion protein thereof and, optionally, other components. For

example, pharmaceutically acceptable carriers suitable for use with a composition of a G protein immunogen or fusion protein thereof, or cocktail of two or more G protein immunogens or fusion proteins thereof, or cocktail of G, F, and M immunogens or fusion proteins thereof, may include, for example, a thickening agent, a buffering agent, a solvent, a humectant, a preservative, a chelating agent, an adjuvant, and the like, and combinations thereof.

Exemplary adjuvants include alum (aluminum hydroxide, REHYDRAGEL®), aluminum phosphate, proteosome adjuvant with LPS (protollin) or without LPS (see, e.g., U.S. Patent Nos. 5,726,292 and 5,985,284, and U.S. Patent

10 Application Publication Nos. 2001/0053368 and US 2003/0044425), virosomes, liposomes with and without Lipid A, Detox (Ribi/Corixa), MF59, or other oil and water emulsions type adjuvants, such as nanoemulsions (see, e.g., U.S. Patent No. 5,716,637) and submicron emulsions (see, e.g., U.S. Patent No. 5,961,970), and Freund's complete and incomplete. Pharmaceutically acceptable carriers for therapeutic use are well known in the pharmaceutical art, and as described herein and, for example, in Remington's Pharmaceutical Sciences, Mack Publishing Co. (A.R. Gennaro, ed., 18th Edition, 1990) and in CRC Handbook of Food, Drug, and Cosmetic Excipients, CRC Press LLC (S.C. Smolinski, ed., 1992).

In certain embodiments, the G protein immunogens and fragments or variants thereof (including fusion proteins and multivalent compositions) are formulated with proteosome. As used herein, "proteosome" or "projuvant" refers to preparations of outer membrane proteins (OMPs, also known as porins) from Gramnegative bacteria, such as Neisseria species (see, e.g., Lowell et al., J. Exp. Med. 167:658, 1988; Lowell et al., Science 240:800, 1988; Lynch et al., Biophys. J. 45:104, 1984; Lowell, in "New Generation Vaccines" 2nd ed., Marcel Dekker, Inc., New York, Basil, Hong Kong, page 193, 1997; U.S. Patent No. 5,726,292; U.S. Patent No. 4,707,543), which are useful as a carrier or an adjuvant for immunogens, such as bacterial or viral antigens. Proteosomes are hydrophobic and safe for human use, and comparable in size to certain viruses. Proteosomes have the interesting ability to auto-assemble into vesicle or vesicle-like OMP clusters of 20-800 nm, and to noncovalently incorporate, coordinate, associate (e.g., electrostatically or hydrophobically), or

otherwise cooperate with protein antigens (Ags), particularly antigens that have a hydrophobic moiety. Any preparation method that results in the outer membrane protein component in vesicular or vesicle-like form, including multi-molecular membranous structures or molten globular-like OMP compositions of one or more OMPs, is included within the definition of Proteosome. Proteosomes may be prepared, for example, as described in the art (see, e.g., U.S. Patent Nos. 5,726,292 or 5,985,284).

In certain embodiments, the G protein immunogens and fragments or variants thereof (including fusion proteins and multivalent compositions) are formulated with protollin. As used herein, "proteosome:LPS" or "protollin" (also known as "IVX-908") refers to preparations of projuvant admixed as described herein with at least one kind of liposaccharide to provide an OMP-LPS composition (which can function as an immunostimulatory composition). Thus, the OMP-LPS adjuvant can be comprised of two of the basic components of Protollin, which include (1) an outer membrane protein preparation of Proteosomes (i.e., projuvant) prepared from Gramnegative bacteria, such as *Neisseria meningitides*, and (2) a preparation of one or more liposaccharides. It is also contemplated that components of Protollin may be or include lipids, glycolipids, glycoproteins, small molecules, or the like. The Protollin may be prepared, for example, as described in U.S. Patent Application Publication No. 2003/0044425.

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20 Projuvant is generally used in conjunction with antigens (naturallyoccurring or modified) that possess a naturally occurring, modified, or supplementary
hydrophobic moiety or portion (also referred to as a "foot" or "anchor"). Protollin
(containing exogenously added LPS) can also be used with an antigen that does not
contain a hydrophobic foot domain and that can be largely hydrophilic in nature.

Protollin can be admixed or combined with an antigen containing a hydrophobic foot, an antigen lacking a hydrophobic foot, or with a combination of antigens having and not having a hydrophobic portion or foot.

As used herein, "liposaccharide" (such as that used in preparing protollin) refers to native (isolated or prepared synthetically with a native structure) or modified lipopolysaccharide or lipooligosaccharide (collectively, also referred to as "LPS") derived from Gram-negative bacteria, such as Shigella flexneri or Plesiomonas

shigelloides, or other Gram-negative bacteria (including Alcaligenes, Bacteroides, Bordetella, Borrellia, Brucella, Campylobacter, Chlamydia, Citrobacter, Edwardsiella, Ehrlicha, Enterobacter, Escherichia, Francisella, Fusobacterium, Gardnerella, Hemophillus, Helicobacter, Klebsiella, Legionella, Leptospira (including Leptospira interrogans), Moraxella, Morganella, Neiserria, Pasteurella, Proteus, Providencia, other Plesiomonas, Porphyromonas (including Porphyromonas gingivalis), Prevotella, Pseudomonas, Rickettsia, Salmonella, Serratia, other Shigella, Spirillum, Veillonella, Vibrio, or Yersinia species). The liposaccharide may be in a detoxified form (i.e., having the Lipid A core removed) or may be in a form that has not been detoxified. In the instant disclosure, the liposaccharide need not be and preferably is not detoxified.

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The two components of an OMP-LPS adjuvant may be formulated at specific initial ratios to optimize interaction between the components resulting in stable association and formulation of the components for use in the preparation of an immunogenic composition of the invention. The process generally involves the mixing of components in a selected detergent solution (e.g., Empigen BB, Triton X-100, or Mega-10) and then effecting complexing of the OMP and LPS components while reducing the amount of detergent to a predetermined, preferred concentration, by dialysis or, preferably, by diafiltration/ultrafiltration methodologies. Mixing, coprecipitation, or lyophilization of the two components may also be used to effect an adequate and stable association or formulation. In a preferred embodiment, an immunogenic composition comprises one or more G protein immunogens and an adjuvant, wherein the adjuvant comprises a Projuvant (i.e., Proteosome) and liposaccharide.

In certain embodiments, the final liposaccharide content by weight as a percentage of the total Proteosome protein can be in a range from about 1% to about 500%, more preferably in range from about 10% to about 200%, or in a range from about 30% to about 150%. Another embodiment includes an adjuvant wherein the Proteosomes are prepared from Neisseria meningitides and the liposaccharide is prepared from Shigella flexneri or Plesiomonas shigelloides, and the final liposaccharide content is between 50% to 150% of the total Proteosome protein by weight. In another embodiment, Proteosomes are prepared with endogenous

lipooligosaccharide (LOS) content ranging from about 0.5% up to about 5% of total OMP. Another embodiment of the instant invention provides Proteosomes with endogenous liposaccharide in a range from about 12% to about 25%, and in a preferred embodiment between about 15% and about 20% of total OMP. The instant disclosure also provides a composition containing liposaccharide derived from any Gram-negative bacterial species, which may be from the same Gram-negative bacterial species that is the source of Proteosomes or is a different bacterial species.

In certain embodiments, the Proteosome or Protollin to G protein immunogen ratio in the immunogenic composition is greater than 1:1, greater than 2:1, greater than 3:1 or greater than 4:1. The ratio can be as high as 8:1 or higher. In other embodiments, the ratio of Proteosome or Protollin to coronavirus antigen of the immunogenic composition ranges from about 1:1 to about 1:500, preferably the ratio is at least 1:5, at least 1:10, at least 1:20, at least 1:50, or at least 1:100. An advantage of Protollin:G protein immunogen ratios ranging from 1:2 to 1:200 is that the amount of Proteosome-based adjuvant can be reduced dramatically with no significant effect on the ability of a G protein immunogen to elicit an immune response.

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As used herein, "pharmaceutically acceptable salt" refers to salts of the compounds of the present invention derived from the combination of such compounds and an organic or inorganic acid (acid addition salts) or an organic or inorganic base (base addition salts). The compounds of the present invention may be used in either the free base or salt forms, with both forms being considered as being within the scope of the present invention.

In addition, the pharmaceutical composition of the instant invention may further include a diluent or excipient, such as water or phosphate buffered saline (PBS).

25 Preferably, a diluent or excipient is PBS with a final phosphate concentration range from about 0.1 mM to about 1 M, more preferably from about 0.5 mM to about 500 mM, even more preferably from about 1 mM to about 50 mM, and most preferably from about 2.5 mM to about 10 mM; and the final salt concentration ranges from about 100 mM to about 200 mM and most preferably from about 125 mM to about 175 mM.

30 Preferably, the final PBS concentration is about 5 mM phosphate and about 150 mM salt (such as NaCl). In certain embodiments, pharmaceutical compositions of the

instant disclosure comprising any of the herein described RSV immunogens or cocktails of RSV immunogens are sterile.

The compositions can be sterile either by preparing them under an aseptic environment or they can be terminally sterilized using methods available in the art. Many pharmaceuticals are manufactured to be sterile and this criterion is defined by the USP XXII <1211>. Sterilization in this embodiment may be accomplished by a number of means accepted in the industry and listed in the USP XXII <1211>, including gas sterilization, ionizing radiation or filtration. Sterilization may be maintained by what is termed aseptic processing, defined also in USP XXII <1211>.

10 Acceptable gases used for gas sterilization include ethylene oxide. Acceptable radiation types used for ionizing radiation methods include gamma, for instance from a cobalt 60 source and electron beam. A typical dose of gamma radiation is 2.5 MRad. When appropriate, filtration may be accomplished using a filter with suitable pore size, for example 0.22 μm and of a suitable material, for instance Teflon[®]. The term "USP" refers to U.S. Pharmacopeia (see www.usp.org; Rockville, MD).

The present description also pertains to methods for treating or preventing RSV infection, comprising administering to a subject in need thereof a composition comprising at least one respiratory syncytial virus G protein immunogen or fragment thereof comprising an amino acid sequence that is at least 80% identical to SEQ ID NO: 2, wherein the G protein immunogen has an epitope that elicits a protective immune response without eliciting an immunopathological response or with a reduced immunopathological response, and pharmaceutically acceptable carrier, diluent, or excipient, at a dose sufficient to elicit an immune response specific for one or more G protein immunogen or fragment thereof. In certain embodiments, an infection is due to a subgroup A, subgroup B, or both subgroups A and B of RSV. In certain preferred embodiments, the G protein immunogens used in any of the compositions and methods described herein have an amino acid sequence as set forth in SEQ ID NO:6, 8, 10, 12, 14,16, 18, 20, 22, 56, 58, 60, 62, 64, 66, 68 or 70, more preferably SEQ ID NO:6, SEQ ID NO:56 or SEQ ID NO:58.

The present description also pertains to methods for reducing the risk of an immunopathological response associated with RSV infection, comprising

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administering to a subject in need thereof a composition comprising at least one respiratory syncytial virus G protein immunogen or fragment thereof comprising an amino acid sequence that is at least 80% identical to SEQ ID NO: 2, wherein the G protein immunogen has an epitope that elicits a protective immune response without eliciting an immunopathological response or with a reduced immunopathological response, and pharmaceutically acceptable carrier, diluent, or excipient, at a dose sufficient to elicit an immune response specific for one or more G protein immunogen or fragment thereof. In certain embodiments, an infection is due to a subgroup A, subgroup B, or both subgroups A and B of RSV. In certain preferred embodiments, the G protein immunogens used in any of the compositions and methods described herein have an amino acid sequence as set forth in SEQ ID NOS:6, 8, 10, 12, 14,16, 18, 20, 22, 56, 58, 60, 62, 64, 66, 68 or 70, more preferably SEQ ID NO:6, SEQ ID NO:56 or SEQ ID NO:58.

A subject suitable for treatment with a RSV immunogen formulation

may be identified by well-established indicators of risk for developing a disease or
well-established hallmarks of an existing disease. For example, indicators of an
infection include fever, pus, microorganism positive cultures, inflammation, and the
like. Infections that may be treated or prevented with a RSV immunogen vaccine of the
subject invention include those caused by or due to RSV, whether the infection is

primary, secondary, opportunistic, or the like. Examples of RSV include any subtype,
strain, antigenic variant, and the like, of these viruses. For preventative purposes, for
example, certain known risk factors for acquiring an RSV infection include premature
birth, children with chronic lung disease, children that attend daycare, presence of
school-age siblings in the home, exposure to passive smoke in the home, and
immunocompromised subjects (adult and children).

Pharmaceutical compositions containing one or more RSV immunogens of the instant description may be in any form that allows the composition to be administered to a subject, such as a human or animal. For example, G protein immunogen, fusion protein, and multivalent compositions of the present description may be prepared and administered as a liquid solution or prepared as a solid form (e.g., lyophilized), which may be administered in solid form, or resuspended in a solution in

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conjunction with administration. The hybrid polypeptide composition is formulated so as to allow the active ingredients contained therein to be bioavailable upon administration of the composition to a subject or patient or bioavailable via slow release. Compositions that will be administered to a subject or patient take the form of one or more dosage units, where for example, a tablet may be a single dosage unit, and a container of one or more compounds of the invention in aerosol form may hold a plurality of dosage units. In certain preferred embodiments, any of the herein described pharmaceutical compositions comprising a RSV immunogen or cocktail of immunogens of the invention are in a container, preferably in a sterile container.

10 In one embodiment, the therapeutic composition is administered nasally, wherein cells, such as cells located in the nasal associated lymphoid tissue, can take up an RSV immunogen or cocktail composition of this disclosure. Other typical routes of administration include, without limitation, enteral, parenteral, transdermal/transmucosal, nasal, and inhalation. The term "enteral", as used herein, is a route of administration in which the immunogenic composition is absorbed through the gastrointestinal tract or oral mucosa, including oral, rectal, and sublingual. The term "parenteral", as used herein, describes administration routes that bypass the gastrointestinal tract, including intraarterial, intradermal, intramuscular, intranasal, intraocular, intraperitoneal, intravenous, subcutaneous, submucosal, and intravaginal injection or infusion techniques. The term "transdermal/transmucosal", as used herein, 20 is a route of administration in which the immunogenic composition is administered through or by way of the skin, including topical. The terms "nasal" and "inhalation" encompass techniques of administration in which an immunogenic composition is introduced into the pulmonary tree, including intrapulmonary or transpulmonary. Preferably, the compositions of the present invention are administered nasally. 25

In another embodiment, the instant compositions comprising at least one respiratory syncytial virus G protein immunogen or fragment thereof can be used in prophylactic methods. For example, an RSV immunogen or cocktail composition of the invention may be administered to a mother during gestation to prevent an RSV infection in the mother and to provide passive immunity to the fetus or new born. A prophylactic method may comprise administering to a first subject a composition

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comprising an RSV immunogen and pharmaceutically acceptable carrier, diluent or excipient, followed by administration to a second subject of a second composition comprising at least one respiratory syncytial virus immunogen wherein said first composition comprises a different RSV immunogen than that administered to the second subject and the second composition comprises at least one respiratory syncytial virus G protein immunogen or fragment thereof comprising an amino acid sequence that is at least 80% identical to SEQ ID NO: 2 and a pharmaceutically acceptable carrier, diluent or excipient, wherein the G protein immunogen has an epitope that elicits a protective immune response without eliciting an, or with a reduced, immunopathological response. In certain embodiments, the G protein immunogens for prophylactic use can have an amino acid sequence as set forth in SEQ ID NO:6, 8, 10, 12, 14,16, 18, 20, 22, 56, 58, 60, 62, 64, 66, 68 or 70, more preferably SEQ ID NO:6, SEQ ID NO:56 or SEQ ID NO:58.

A representative first subject can be a mother during gestation and a representative second subject can be the mother's newborn child. Each composition is provided at a dose sufficient to elicit an immune response specific for one or more RSV immunogen (such as G protein immunogens described herein). For instance, not wishing to be bound by theory, G protein immunogens and compositions thereof can be administered systemically (e.g., intravenously) to the mother, which would elicit IgG antibodies similar to the antibodies the mother already has due to exposure to RSV. 20 The newborn child can then be immunized via the mucosa (e.g., intranasally), which would elicit secretory IgA antibodies - hence, the G protein immunogens administered via the mucosa will not be detected by the systemic maternal (IgG) antibodies the child inherited because the IgG antibodies will not be at the mucosal interface. That is, the maternally inherited antibodies will not adversely affect the IgA response elicited by 25 intranasal immunization of the child. In certain embodiments, the administered compositions may prevent an infection due to a subgroup A, subgroup B, or both subgroups A and B of RSV. A subject suitable for treatment with a RSV immunogen formulation may be identified by well-established indicators of risk for developing a disease or well-established hallmarks of an existing disease as described herein and is 30 known in the art. Infections that may be treated with a RSV immunogen of the subject

invention include those caused by or due to RSV, whether the infection is primary, secondary, opportunistic, or the like. Examples of RSV include any strain, subtype, antigenic variant, and the like of these viruses.

The invention further provides a plurality of antibodies produced by the method for preventing a RSV infection that comprises administering to a subject a composition of the subject invention at a dose sufficient to elicit antibodies specific for one or more RSV immunogen wherein said G protein immunogen has an epitope that elicits a protective immune response without eliciting an, or with a reduced, immunopathological response. In one embodiment, the antibodies comprise at least one antibody specific for a subgroup A RSV, or a subgroup B RSV, or for both subgroup A and B RSVs. In another embodiment, a method for treating or preventing a RSV infection comprises administering to a subject a composition comprising a pharmaceutically acceptable carrier, with or without an adjuvant, and a plurality of antibodies of the subject invention.

In addition, a subject at risk for an RSV infection can have a plurality of antibodies according to this description administered before, simultaneous with, or after administration of a composition comprising at least one different respiratory syncytial virus G protein immunogen or fragment thereof comprising an amino acid sequence that is at least 80% identical to SEQ ID NO: 2 and pharmaceutically acceptable carrier, diluent or excipient, according to the instant description. In certain preferred embodiments, the G protein immunogens used in any of the compositions and methods described herein have an amino acid sequence as set forth in SEQ ID NOS:6, 8, 10, 12, 14,16, 18, 20, 22, 56, 58, 60, 62, 64, 66, 68 or 70, more preferably SEQ ID NO:6, SEQ ID NO:56 or SEQ ID NO:58. In some embodiments, antibodies specific for one or more RSV immunogens can be provided passively, while the subject is vaccinated to actively elicit antibodies against one or more different RSV immunogens.

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In another aspect, the RSV G protein immunogens and fragments, variants thereof of the present invention are utilized to elicit antibodies specific for at least one epitope present on the G protein immunogens and fragments or variants thereof provided herein. Accordingly, the present invention also provides such antibodies. In preferred embodiments the antibodies bind to specific protective epitopes

present on an RSV G protein. Within the context of the present invention, the term "antibodies" includes polyclonal antibodies, monospecific antibodies, monoclonal antibodies, anti-idiotypic antibodies, fragments thereof such as F(ab')2 and Fab fragments, and recombinantly or synthetically produced antibodies. Such antibodies incorporate the variable regions that permit a monoclonal antibody to specifically bind, which means an antibody is able to selectively bind to a peptide or polypeptide from an RSV G protein from subtype A or B. "Specific for" refers to the ability of a protein (e.g., an antibody) to selectively bind a polypeptide or peptide encoded by a nucleic acid molecule encoding a from an RSV G protein from subtype A or B, or a synthesized RSV G protein from subtype A or B, of this invention. Association or "binding" of an antibody to a specific antigen generally involve electrostatic interactions, hydrogen bonding, Van der Waals interactions, and hydrophobic interactions. Any one of these or any combination thereof can play a role in the binding between an antibody and its antigen. Such an antibody generally associates with an antigen, such as a G protein immunogen, with an affinity constant (K_a) of at least 10⁴, preferably at least 10⁵, more preferably at least 10⁶, still more preferably at least 10⁷ and most preferably at least 10⁸. Affinity constants may be determined by one of ordinary skill in the art using wellknown techniques (see Scatchard, Ann. N.Y. Acad. Sci. 51:660-672, 1949). The affinity of a monoclonal antibody or antibody can be readily determined by one of ordinary skill in the art (see Scatchard, Ann. N.Y. Acad. Sci. 51:660-672, 1949).

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In addition, the term "antibody," as used herein, includes naturally occurring antibodies as well as non-naturally occurring antibodies, including, for example, single chain antibodies, chimeric, bifunctional and humanized antibodies, fully human antibodies, as well as antigen-binding fragments thereof. Such non-naturally occurring antibodies may be constructed using solid phase peptide synthesis, may be produced recombinantly, or may be obtained, for example, by screening combinatorial libraries consisting of variable heavy chains and variable light chains (Huse et al., Science 246:1275-1281 (1989)). These and other methods of making, for example, chimeric, humanized, CDR-grafted, single chain, and bifunctional antibodies are well known in the art (Winter and Harris, Immunol. Today 14:243, 1993; Ward et al., Nature 341:544, 1989; Harlow and Lane, Antibodies: A Laboratory Manual, Cold

Spring Harbor Laboratory, New York, 1992; Borrabeck, Antibody Engineering, 2d ed., Oxford Univ. Press, 1995; Hilyard et al., Protein Engineering: A practical approach, IRL Press, 1992).

Polyclonal antibodies can be readily generated by one of ordinary skill in

the art from a variety of warm-blooded animals, including horses, cows, goats, sheep, dogs, chickens, turkeys, rabbits, mice, or rats. Briefly, the desired G protein immunogen or fragment thereof, or mixtures of RSV immunogens, or variants thereof are administered to immunize an animal through parenteral, intraperitoneal, intramuscular, intraocular, or subcutaneous injections. The immunogenicity of the hybrid polypeptide of interest may be increased through the use of an adjuvant, such as alum and Freund's complete or incomplete adjuvant. Following several booster immunizations over a period of weeks, small samples of serum are collected and tested for reactivity to the desired immunogen. Once the titer of the animal has reached a plateau in terms of its reactivity to a G protein immunogen of the invention, larger quantities of polyclonal immune sera may be readily obtained either by weekly bleedings or by exsanguinating the animal.

The RSV immunogens of the instant invention can be easily identified using in vitro and in vivo assays known in the art and as described herein.

Representative assays are described in the Examples. Similarly, several assays are available as described herein to examine the activity of the antibodies elicited by the RSV immunogens of the subject invention.

All of the U.S. patents, U.S. patent application publications, U.S. patent applications, foreign patents, foreign patent applications and non-patent publications referred to in this specification or listed in the Application Data Sheet, are incorporated herein by reference, in their entirety. The invention having been described, the following examples are intended to illustrate, and not limit, the invention.

EXAMPLES

EXAMPLE 1

PREPARATION OF PROTEOSOMES

Immunogens of the instant invention may be formulated with proteosomes by way of non-covalent interactions to form a vaccine composition capable of eliciting a protective immune response in an immunized human or animal subject. Proteosomes of the instant application are mucosal adjuvant delivery vehicles comprising outer membrane proteins purified from, for example, Group B type 2 Neisseria meningitides. The use of proteosomes for the formulation of vaccines has been reviewed by Lowell, G. H., in "New Generation Vaccines 2nd ed., Marcel Dekker, 10 Inc., New York, Basil, Hong Kong (1997) pages 193-206. Proteosomes of the instant invention may be prepared by extraction of phenol-killed bacterial paste with a solution of 6% Empigen BB (EBB) (Albright and Wilson, Whithaven, UK) in 1 M calcium chloride followed by precipitation with ethanol, solubilization in 1% EBB-Tris/EDTA-15 saline and then precipitation with ammonium sulfate. The precipitates are re-solubilized in the 1% EBB buffer, dialyzed, and stored in 0.1% EBB at -70°C. Alternative processes may be used in the preparation of proteosomes, for example, proteosomes may be prepared by omitting the ammonium sulfate precipitation step to shorten the process. Preparation of proteosomes are disclosed in U.S. Patent Application 20 Publication No. 2001/0053368 and in U.S. Patent No. 6,476,201 B1.

EXAMPLE 2

PREPARATION OF LIPOSOMES

Immunogens of the instant invention may be combined non-covalently with liposomes as a vaccine composition capable of eliciting a protective immune response in an immunized human or animal subject. Immunogens may be encapsulated with multilamellar liposomes according to procedures known to those of ordinary skill in the art using, for example, a dehydration coupled reconstitution method (Kirby and Gregoriadis, *BioTechnology* 2:979, 1984). Briefly, liposomes are prepared by

sonication of dioleoylphosphatidyl choline (DOPC/cholesterol, Sigma Chemical Co., St. Louis, MO; 5:1, W/W) at a final lipid concentration of 30 mg/ml in PBS or generating liposomes using *Deinococcus radiodurans* lipids or α-galactosylphosphotidylglycerolalkylamine as desribed in Huang and Anderson, *Vaccine* 20:1586, 2002, in the presence or absence of antigen. The liposome, with or without one or more immunogens, are lyophilized and resuspended in sterile water. Immunogen that is not incorporated into liposomes may be removed by repeated washing and centrigugation (*e.g.*, microcentrifugation for 1 min at 13,200 rpm) of the liposomes in phosphate buffered saline (PBS). The protein content of washed liposomes, with and without immunogen, is determined by, for example, quantitative silver-stained SDS-PAGE using calibrated amounts known protein standards, such as serum albumin. The protein content of the liposomes is determined and adjusted as desired, for example, the protein content may be adjusted to 0.3 mg per mg of lipid (as liposomes) per ml.

In some cases, to evaluate the manner in which the protein antigen interacts with a liposome, liposomes containing aliquots of G protein immunogens and fragments thereof (wild type or mutant) may be incubated for 1 hour at 37°C in PBS with proteinase K (Gibco/BRL) at 1.0, 0.1 and 0.01 µg/ml. Some incubations may also contain 1% Triton X-100 to disrupt liposomes, thereby allowing complete access of the proteinase K to the proteins, fusion proteins, or polypeptide fragments thereof. Incubations are terminated and samples analyzed by SDS-PAGE and silver staining. Such procedures may be used to determine the extent of liposome encapsulation of the immunogen (e.g., one or more viral proteins) preparation.

EXAMPLE 3

25 PREPARATION OF NUCLEIC ACIDS AND EXPRESSION CONSTRUCTS
ENCODING G PROTEIN IMMUNOGENS AND FRAGMENTS THEREOF

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G protein encoding nucleic acid sequences from the RSV (Long strain) corresponding to amino acids 128-229, as well as, for example, mutant 128-229 sequences were amplified from viral RNA by RT-PCR, and the resultant PCR products

cloned into the EcoRI and XhoI sites of a pET-32-LIC bacterial expression plasmid (Novagen, Madison, WI). Site-directed mutagenesis of the RSV G128-229 protein sequence was performed according to the Stratagene QuikChange® site-directed mutagenesis protocol. Briefly, PCR was performed on template pET-32-LIC-G128-229

5 DNA (G128-229 sequence cloned into EcoRI and XhoI sites).

In these experiments, the primer pairs designed for mutagenesis were as follows:

CCTGCTGGGCTGCCAAAAGAATACCAAACAAAAAACCAGG (SEQ ID NO:37) and CCTGGTTTTTGTTTGGTATTCTTTTGCAGGCAGCCCAGC AGG (SEQ ID NO:38) (for the G128-229, I185A mutant);

CTGCTGGGCTATCGCCAAAAGAATACCAAACAAAAAACCAGG (SEQ ID NO:39) and CCTGGTTTTTTGTTTGGTATTCTTTTGGCGATAGCCCAGCAG (SEQ ID NO:40) (for the G128-229, C186A mutant);

CTGCTGGGCTATCTGCGCAAGAATACCAAACAAAAAACCAGG (SEQ ID NO:41) and CCTGGTTTTTTGTTTGGTATTCTTGCGCAGATAGCCCAGCAG (SEQ ID NO:42) (for the G128-229, K187A mutant);

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CTGCTGGGCTATCTGCAAAGCAATACCAAACAAAAAACCAGG (SEQ ID NO:43)
and CCTGGTTTTTTGTTTGGTATTGCTTTGCAGATAGCCCAGCAG (SEQ ID NO:44) (for the G128-229, R188A mutant);

25 CTGCTGGGCTATCTGCAAAAGAGCACCAAACAAAAAACCAGG (SEQ ID NO:45) and CCTGGTTTTTTGTTTGGTGCTCTTTTGCAGATAGCCCAGCAG (SEQ ID NO:46) (for the G128-229, I189A mutant);

CTGCTGGGCTATCTGCAAAAGAATAGCAAACAAAAAACCAGG (SEQ ID NO:47)
and CCTGGTTTTTTGCTATTCTTTTGCAGATAGCCCAGCAG (SEQ ID NO:48)
(for the G128-229, P190A mutant);

CTGCAAAAGAATACCAGCCAAAAAACCAGGAAAGAAAACCACC (SEQ ID NO:49) and GGTGGTTTTCCTTGGTTTTTTTGGCTGGTATTCTTTTGCAG (SEQ ID NO:50) (for the G128-229, N191A mutant);

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- CTGGGCTATCTGCAAAAGAATACCAAACGCAAAACCAGGAAAG (SEQ ID NO:51) and CTTTCCTGGTTTTGCGTTTTGGTATTCTTTTGCAGATAGCCCAG (SEQ ID NO:52) (for the G128-229, K192A mutant);
- 10 GCAAAAGAATACCAAACAAAGCACCAGGAAAGAAAACCACCAC (SEQ ID NO:53) and GTGGTGGTTTTCTTTCCTGGTGCTTTGTTTGGTATTCTTTTGC (SEQ ID NO:54) (for the G128-229, K193A mutant).

Thioredoxin (Trx)-fusion proteins containing wild type, and the above mutant RSV G protein fragments were prepared as described in Example 4.

EXAMPLE 4

PRODUCTION OF G PROTEIN IMMUNOGENS AND FRAGMENTS THEREOF

RSV G protein immunogens can be prepared as pharmaceutical compositions by mixing with a pharmaceutically acceptable carrier, excipient or

20 diluent. For example, the RSV G protein sequences described herein and encoded by nucleic acid contained in modified pET-32-LIC plasmids were expressed as thioredoxin (Trx)-fusion proteins in transformed *E. coli* BL21/DE3 cells following induction with IPTG. All Trx-fusion proteins were recovered from transformed cell pellets by extraction with 8M urea, followed by affinity purification using TALON® (Clontech,

25 Palo Alto, CA) and dialysis against PBS. Purified Trx-G128-229 polypeptides were freed from contaminating endotoxin by treatment with polymyxin B beads (BioRad, Mississauga, ON, Canada). Details and modifications of this procedure are well known to those of ordinary skill in the art. Upon use for immunization, immunogens can be

further combined or admixed with an adjuvant, such as alum or a proteosome-based adjuvant.

EXAMPLE 5

PREPARATION OF RSV

5 RSV (Long strain) was obtained from the American Type Culture Collection and propagated on HEp-2 cells cultured in Earle's Minimal Essential Medium (MEM) containing penicillin G (100 U/ml) and streptomycin sulfate (100 ug/ml) and supplemented with 1% serum (fetal calf serum/calf serum, 1:3). Cells and virus were verified negative for mycoplama contamination by PCR assay 10 (American Type Culture Collection). As described in this Example, confluent monolayer cultures of HEp-2 cells were inoculated with RSV (Long strain) at a multiplicity of infection (MOI) of 1, adsorbed 90 min at 4°C, washed and incubated at 37°C in RPMI-1640 medium (Sigma, St. Louis, MO) supplemented with 1% fetal calf serum (Sigma, St. Louis, MO). Cultures were harvested after 24-30h, at which time the 15 cell monolayers were almost completely fused; virus was released from cells by disruption with a hand-held Teflon scraper (Gibco-BRL) and cellular debris was removed by microcentrifugation for 5 min at 13,000 x g. The supernatant was used as the source of virus for mouse challenge studies (see Example 6).

EXAMPLE 6

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MOUSE IMMUNIZATION

All immunizations were performed using BALB/c mice (Charles River, Ste. Constance, QC, Canada), which were anaesthetized with ketamine (2.3 mg/mouse; Bimeda-MTC Pharmaceuticals, Cambridge, ON, Canada) and xylazine (0.5 mg/mouse; Bayer, Toronto, ON, Canada). For vaccine/challenge experiments, groups of seven to nine BALB/c mice (6-8 wks old) were immunized twice subcutaneously, at 14-day intervals, with PBS/alum, Trx-G128-229, or mutant Trx-G128-229 proteins, in PBS/alum (10 µg protein in a volume of 50 µl). Fourteen days after the second dose, mice were challenged intranasally with RSV (2 x 10⁶ pfu in 50 µl). Mice were

sacrificed using sodium pentobarbital four days later and assayed for lung virus titers and leukocyte infiltration in bronchoalveolar fluids according to procedures well known to a person of ordinary skill in the art.

Immunoblot analysis demonstrated that serum antibodies raised against amino acids 128-229 of RSV G protein were capable of specifically recognizing RSV G protein in mice immunized with wild type or mutant Trx-G128-229 proteins (Fig. 1). Extracts of RSV-infected HEp-2 cells were resolved by SDS-PAGE, and transferred to membranes (e.g., polyvinyldene difluoride (PVDF) membranes). Membranes containing transferred protein were blocked (to prevent non-specific interactions) with 10 4% skim milk and 0.5% casein (Hammerstein grade) in TBST (0.8% NaCl, 0.1% Tween-20, 20mM Tris, pH 7.6) by overnight incubation at room temperature. Blocked membranes were then incubated with serum samples, washed with TBST, followed by 1 hour incubation with horse-radish peroxidase (HRP)-conjugated goat antimouse antibody, and then signal was then detected using diaminobezidine (DAB; 1 mg/ml, 0.03% NiCl2 and 0.1% H2O2, according to procedures known in the art. A strong G 15 protein-specific antibody (IgG) response was observed with wild type and N191A mutant proteins. Very little RSV G protein antibody specific signal was observed in sera obtained from mice immunized with I185A or K187A mutant RSV G polypeptide fusion proteins. The remaining Trx-G128-229 mutant proteins induced intermediate levels of RSV G-specific antibodies (Fig. 1). 20

EXAMPLE 7

RSV CHALLENGE OF IMMUNIZED MICE

In these experiments, mice were immunized (as described in Example 6) with either wild type Trx-G128-229 or one of each of the 9 mutants and then challenged with RSV. Induction of eosinophilia was determined according to procedures described in Mader et al. Vaccine 18:1110, 2000. As shown in Figure 2A, wild type Trx-G128-229 and various single mutants protected mice against RSV challenge to varying degrees. Comparatively, the N191A mutated Trx-G128-229 provided better protection than did mutants P190A, R188A and I189A. The remaining Trx-G128-229 mutant

proteins conferred intermediate levels of protection. Furthermore, comparatively, the R188A and N191A mutants demonstrated the highest level of protection. This surprising result indicates that a single point mutation in a G protein can result in a polypeptide capable of eliciting a protective immune response concomitantly with a much reduced immunopathological response (e.g., pulmonary eosinophilia).

EXAMPLE 8 RSV NEUTRALIZATION ASSAY

In these experiments, aliquots of pre-titered RSV were mixed with serially diluted samples of individual mouse sera and incubated for 1 hr at room 10 temperature. Serum from individual mice was collected 14 days after the second of two subcutaneous administrations of an immunogen in alum, as described in Example 6. Sera were assayed for RSV neutralizing antibodies by plaque reduction assay. Mixtures were applied in duplicate to 24-well plates containing 60-80% confluent monolayers of HEp-2 cells, adsorbed for 90 minutes at 4°C, followed by washing and incubation of the plates for 40 h at 37°C in 1 ml of RPMI medium supplemented with 1% fetal calf 15 serum. After incubation, the monolayers were fixed with 15% formaldehyde and stained with 0.01% crystal violet for visualization of viral plaques. Plaque reduction is calculated as the plaque reduction neutralization titer₅₀ (PRNT₅₀), which is the reciprocal dilution of sera required to neutralize 50% of RSV plaques on a subconfluent monolayer of HEp-2 cells.

Results of an RSV plague reduction assay are shown in Table 1. The RSV neutralization titers in sera from immunized mice showed a strong dependence of neutralizing antibody responses upon the amino acid sequence within the 185-193 region of the Trx-G128-229 protein used for immunization (similar to the immunization results of Example 6).

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Table 1. Neutralization titers of sera from mice immunized with Trx-G variant proteins

	Immunogen	PRNT ₅₀ *
	PBS	7 ± 3
	Trx-G128-229	144 ± 37
5	Trx-G128-229 (I185A)	18 ± 5
	Trx-G128-229 (C186A)	81 ± 12
	Trx-G128-229 (K187A)	34 ± 11
	Trx-G128-229 (R188A)	39 ± 13
	Trx-G128-229 (I189A)	115 ± 38
10	Trx-G128-229 (P190A)	98 ± 26
	Trx-G128-229 (N191A)	95 ± 21
	Trx-G128-229 (K192A)	27 ± 7
	Trx-G128-229 (K193A)	31 ± 12

^{*} PRNT₅₀ (Plaque Reduction Neutralization Titer₅₀) is calculated by determining the reciprocal dilution of sera required to neutralize 50% of RSV plaques on HEp-2 cells. The results are expressed as a mean ± SD.

EXAMPLE 9

RESPONSE OF CYTOKINE MRNA TO RSV G PROTEIN VARIANTS

20 Cytokine mRNA levels in lungs of mice immunized with various RSV G protein variants were measured using a ribonuclease protection assay, which can be an indicator of whether a harmful eosinophilic response will result. One lobe from each mouse lung was stored at -20°C in RNAlaterTM solution (Qiagen, Mississauga, ON, Canada) and subsequently processed for RNA extraction using the RNeasy[®] mini kit

25 (Qiagen, Mississauga, ON, Canada). RNA was quantitated and subjected to

ribonuclease protection assay (RPA) using a transcription kit (BD-Pharmingen, Mississauga, ON, Canada) to synthesize probe from a cytokine (MCK-1) template (BD-Pharmingen, Mississauga, ON, Canada), radiolabeled using α -³²P [UTP] and followed by hybridization and RNase digestion using an RPAIII kit (Ambion, Austin, TX).

5 Reaction mixtures were resolved on a 5% polyacrylamide 8M urea gel according to the manufacturer's instructions followed by drying and autoradiography at -70°C using an intensifying screen.

The RPA results illustrated striking differences among the mice immunized with wild-type or mutant Trx-G proteins and subsequently challenged with RSV (Figure 4). As shown in Figure 4, the Th2 type cytokines most prone to upregulation were IL-4, IL-10, IL-13 and, to a lesser extent, IL-5. G protein variants K193A, P190A and I189A were found to provoke dramatic IL-4, IL-10 and IL-13 responses. Weak IL-4, IL-10 and IL-13 responses were observed with other G protein variants, such as N192A, N191A, R188A, K187A and C186A. The results of the present study highlight the apparent importance of IL-13, which has been recently implicated in asthma (Grunig et al., Science 282: 2261, 1998) as well as in RSV vaccine-induced disease (Johnson and Graham, J. Virol. 73:8485, 1999). High levels of IL-13 and IL-10 correlated well with high levels of eosinophilia observed in RSV-challenged mice that had been immunized with wild type or mutants I189A, P190A, K192A and K193A.

In contrast, mutants such as N191A, K187A and R188A were poor inducers of IL-13, IL-10 and eosinophilia, despite being decent inducers of IL-4.

In comparison, the prototype Th1 cytokine, IFN-γ, was elevated in all experimental mouse groups. By way of example and not wishing to be bound by theory, this may reflect the expression of IFN-γ from NK cells as well as Th1 cells (Trinchieri, Adv. Immunol. 47:187, 1989), the rapid induction of IFN-γ upon RSV infection (Hussell and Openshaw, J. Gen. Virol. 79: 2593, 1998), and/or the prevalent nature of IFN-γ expression even in immune processes in which a Th2 response appears to predominate (Waris et al., J. Virol. 70:2852, 1996; Spender et al., J. Gen. Virol. 79: 1751, 1998; Srikiatkhachorn and Braciale, J. Virol. 71: 678, 1997).

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EXAMPLE 10

PREPARATION OF PROTEOSOMES CONTAINING RSV G PROTEIN IMMUNOGENS

Portions of stock RSV G protein product immunogens (e.g., wild type or mutant peptides) may be formulated with proteosomes using, by way of example, diafiltration/ultrafiltration methods or by using dialysis. For either method, the RSV G protein product is dissolved in, for example, a saline buffered solution containing the desired detergent (e.g., Empigen BB (EBB) at 1% or, at 0.1%-2% of EBB or other suitable detergent depending on the type of detergent used) and is then mixed with proteosomes in saline buffered 1% Empigen solution (or other appropriate detergent at appropriate concentrations) at various proteosome: RSV G (wt/wt) ratios ranging from 1:4 to 8:1, including 1:4, 1:1, 2:1, 4:1 and 8:1. To remove Empigen, the mixture may then be subjected to ultrafiltration/diafilt-ration technology or is exhaustively dialyzed across a dialysis membrane with, for example, a 10,000 molecular weight cut-off (MWCO) or functionally similar membranes with MWCO ranges of 1,000-30,000 against buffered saline for 1-2 weeks at 4°C exchanging at least 500 parts buffer each day. At various steps, immunological assays such as ELISA and single radial immunodiffusion (SRID) may be used to measure potency. The halo immunodiffusion technique is used to determine the content of formulate RSV G antigen with proteosomes at various ratios (for details on the preparation of proteosomes, see, e.g., U.S. Patent Application Publication No. 2001/0053368).

Multivalent vaccines may also be prepared by making individual monovalent proteosome vaccines and then combining them at the required proportions prior to final formulation and fill. Multivalent preparations may also be formulated by pooling individual RSV G antigens in the desired proportions and formulating the mixture with proteosomes. Multivalent vaccine preparations may contain one or more RSV F protein immunogens and/or one or more M protein immunogens in combination with one or more RSV G protein immunogens. The vaccine composition is then passed through membrane filters of 0.8 μm pore size and stored at 4°C prior to and during immunizations.

RSV G protein immunogens (e.g., wild type or mutant peptides in any of the previous forms) may also be formulated with various amounts of proteosome-LPS adjuvant as disclosed in, for example, U.S. Patent No. 6,476,201 B1, and described herein.

EXAMPLE 11

5 IMMUNIZATION WITH PROTOLLIN FORMULATED RSV G PROTEIN IMMUNOGENS

Mice were immunized intranasally with RSV G wild-type (amino acids 128-229) or the mutant (N191A) proteins formulated with Protollin to determine whether RSV-specific systemic and mucosal titers were elicited. BALB/c mice were immunized three times with a dose of 6 μg or 2 μg of either the Trx-(polyHis)-G(128-229) fusion proteins alone, or adjuvanted with protollin or alum. Protollin alone or fusion proteins formulated with protollin were administered intranasally, and alum alone or fusion proteins formulated with alum were administered subcutaneously. Blood was drawn from the saphenous vein after the second dose (day 35) and serum was obtained by exsanguination two weeks after the third dose (day 62).

15 Bronchoalveolar lavage (BAL) samples were also collected on day 62.

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RSV G-specific serum IgG and BAL IgA titers were determined by ELISA. A 10-fold increase in serum IgG titers was observed in mice immunized intranasally with Trx-(polyHis)-G(128-229) formulated with Protollin compared to mice immunized with Trx-(polyHis)-G(128-229) alone, at both the 6 and 2µg doses (Figure 5). There was no significant difference in serum IgG titers between the groups immunized intranasally with Protollin formulations or subcutaneously with alum formulations. Comparable titers were obtained for both G(128-229) wild-type and G(128-229) mutant N191A when given with either adjuvant. RSV G(128-229)-specific BAL IgA titers were significantly higher in the groups having received the G(128-229) immunogens (wild-type or mutant) formulated with Protollin compared to the groups immunized with the immunogens alone (Figure 6). Again, comparable titers were obtained for both wild-type and mutant G(128-229) immunogens. As expected, no IgA was detected in the groups immunized subcutaneously with the G(128-229) immunogens formulated with alum. These results indicate the Protollin formulated

G(128-229) immunogens (wild-type or mutant) vaccines are well tolerated and are immunogenic when administered intranasally.

From the foregoing it will be appreciated that, although specific embodiments of the invention have been described herein for purposes of illustration, various modifications may be made without deviating from the spirit and scope of the invention. Accordingly, the invention is not limited except as by the appended claims.

CLAIMS

1. A method for treating or preventing a respiratory syncytial virus infection, comprising administering to a subject in need thereof a composition comprising at least one respiratory syncytial virus G protein immunogen or fragment thereof comprising an amino acid sequence that is at least 80% identical to SEQ ID NO:2, wherein said G protein immunogen has an epitope that elicits a protective immune response without eliciting an immunopathological response or eliciting a reduced immunopathological response, and a pharmaceutically acceptable carrier, diluent or excipient at a dose sufficient to elicit an immune response specific for one or more G protein immunogen or fragment thereof.

- 2. The method according to claim 1 wherein said G protein immunogen is an amino acid sequence consisting of SEQ ID NO:2.
- 3. The method according to claim 1 wherein said G protein immunogen comprises an amino acid sequence selected from SEQ ID NOS:6, 8, 10, 12, 14,16, 18, 20, 22, 56, 58, 60, 62, 64, 66, 68 or 70.
- 4. The method according to claim 1 wherein said G protein immunogen comprises an amino acid sequence selected from SEQ ID NO:6, SEQ ID NO:56 or SEQ ID NO:58.
- 5. The method according to any one of claims 1 to 4 wherein said G protein immunogen further comprises a hydrophobic moiety.
- 6. The method according to claim 5 wherein said hydrophobic moiety comprises an amino acid sequence.
- 7. The method according to claim 5 wherein said hydrophobic portion is a lipid.

8. The method according to claim 5 wherein said hydrophobic moiety is at the amino-terminus of the fusion protein.

- 9. The method according to claim 5 wherein said hydrophobic moiety is at the carboxy-terminus of the fusion protein.
- 10. The method according to claim 1 wherein the pharmaceutically acceptable carrier, diluent or excipient is a liposome.
- 11. The method according to claim 1 wherein said composition further comprises an adjuvant.
- 12. The method according to claim 11 wherein said adjuvant is alum, protollin, or proteosome.
- 13. The method according to claim 1 wherein said composition further comprises at least one respiratory syncytial virus F protein immunogen or M protein immunogen, wherein said F protein immunogen and said M protein immunogen each has an epitope that elicits a protective immune response without eliciting an immunopathological response or eliciting a reduced immunopathological response.
- 14. The method according to claim 1 wherein said composition has at least two G protein immunogens.
- 15. The method according to any one of claims 1 to 4 wherein said G protein immunogen or fragment thereof further comprises a second amino acid sequence to form a fusion protein.
- 16. The method according to claim 15 wherein said second amino acid sequence is a tag or an enzyme.

17. The method according to claim 15 wherein said second amino acid sequence is thioredoxin, polyhistidine, or a combination thereof.

- 18. The method according to claim 15 wherein said fusion protein further comprises a hydrophobic moiety.
- 19. The method according to claim 18 wherein said hydrophobic moiety is at the amino-terminus of the fusion protein.
- 20. The method according to claim 18 wherein said hydrophobic moiety is at the carboxy-terminus of the fusion protein.
- 21. The method according to claim 17 wherein said fusion protein further comprises a hydrophobic moiety.
- 22. The method according to claim 1 wherein said immunopathological response is eosinophilia or asthma.
- 23. The method according to claim 22 wherein said eosinophilia is pulmonary eosinophilia.
- 24. The method according to claim 1 wherein the infection is due to a subgroup A respiratory syncytial virus.
- 25. The method according to claim 1 wherein the infection is due to a subgroup B respiratory syncytial virus.
- 26. The method according to claim 1 wherein the infection is due to both a subgroup A and a subgroup B respiratory syncytial virus.

27. The method according to any one of claims 1 to 4, 11 and 12 wherein said composition is administered by a route selected from the group consisting of enteral, parenteral, transdermal, transmucosal, nasal, and inhalation.

- 28. The method according to any one of claims 1 to 4, 11 and 12 wherein said composition is administered nasally.
- 29. A plurality of antibodies produced by a method according to any one of claims 1 to 4, 11 and 12.
- 30. A method for treating or preventing a respiratory syncytial virus infection, comprising administering to a subject in need thereof a composition comprising a pharmaceutically acceptable carrier, diluent or excipient, and a plurality of antibodies according to claim 29.
- 31. A composition comprising a respiratory syncytial virus G protein immunogen formulated with a proteosome or protollin, wherein said G protein immunogen comprises an amino acid sequence having at least 80% identity with the sequence set forth in SEQ ID NO:2 or fragment thereof, and wherein said G protein immunogen or fragment thereof has an epitope that elicits a protective immune response without eliciting an immunopathological response or with a reduced immunopathological response.
- 32. The composition according to claim 31 wherein said G protein immunogen is an amino acid sequence consisting of SEQ ID NO:2.
- 33. The composition according to claim 31 wherein said G protein immunogen comprises an amino acid sequence selected from SEQ ID NOS:6, 8, 10, 12, 14,16, 18, 20, 22, 56, 58, 60, 62, 64, 66, 68 or 70.

34. The composition according to claim 31 wherein said G protein immunogen comprises an amino acid sequence selected from SEQ ID NO:6, SEQ ID NO:56 or SEQ ID NO:58.

- 35. The composition according to any one of claims 31 wherein said G protein immunogen further comprises a hydrophobic moiety.
- 36. The composition according to claim 35 wherein said hydrophobic moiety comprises an amino acid sequence.
- 37. The composition according to claim 35 wherein said hydrophobic portion is a lipid.
- 38. The composition according to claim 35 wherein said hydrophobic moiety is at the amino-terminus of the fusion protein.
- 39. The composition according to claim 35 wherein said hydrophobic moiety is at the carboxy-terminus of the fusion protein.
- 40. The composition according to any one of claims 31 to 35 wherein said G protein immunogen or fragment thereof further comprises a second amino acid sequence to form a fusion protein.
- 41. The composition according to claim 40 wherein said second amino acid sequence is a tag or an enzyme.
- 42. The composition according to claim 40 wherein said second amino acid sequence is thioredoxin, polyhistidine, or a combination thereof.
- 43. The composition according to claim 40 wherein said fusion protein further comprises a hydrophobic moiety.

44. The composition according to claim 43 wherein said hydrophobic moiety is at the amino-terminus of the fusion protein.

45. The composition according to claim 43 wherein said hydrophobic moiety is at the carboxy-terminus of the fusion protein.

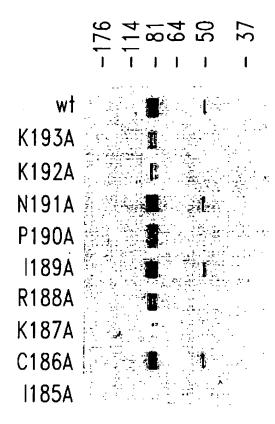
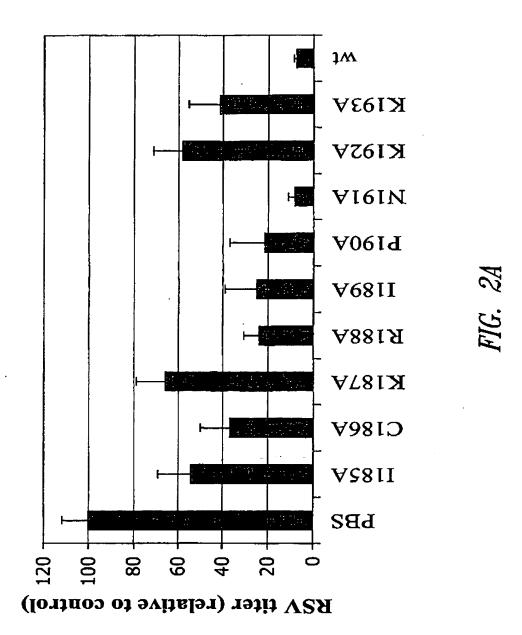
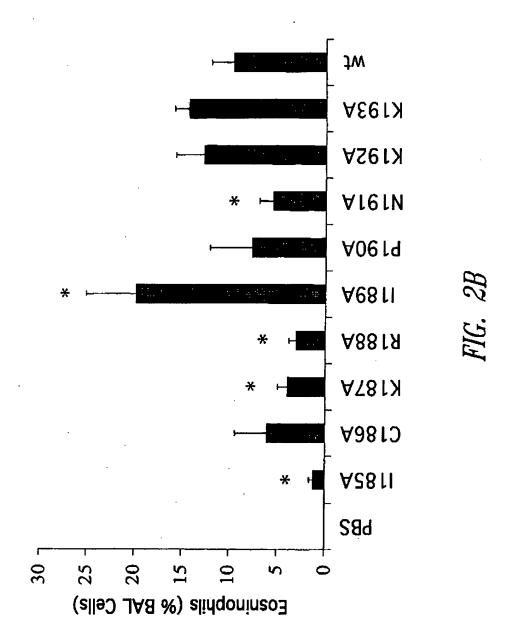


FIG. 1



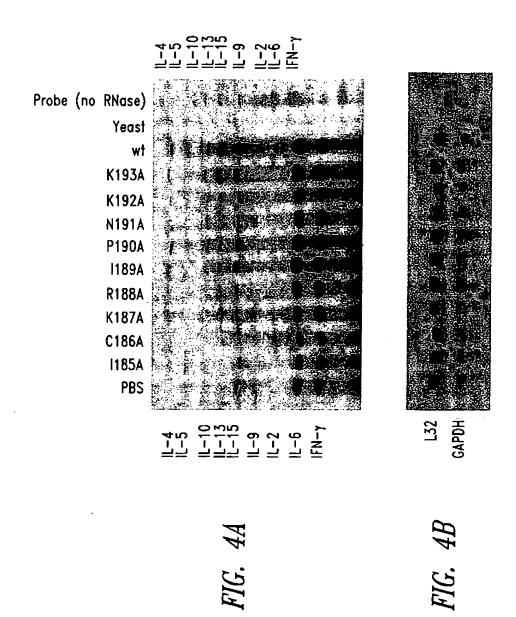


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5/8

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d35 =□ d62 =⊠

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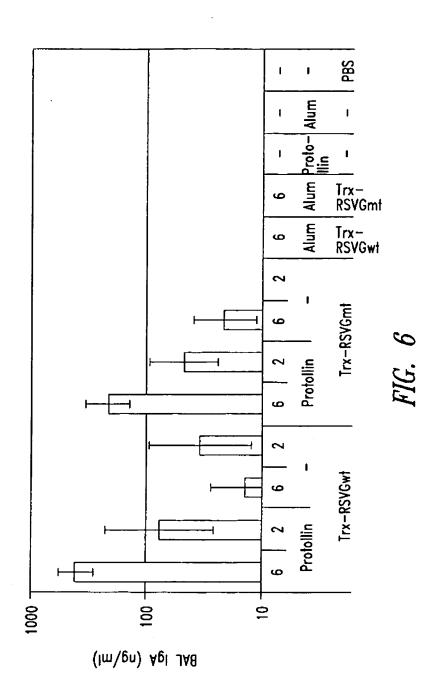
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                                 250
              245
Lys Leu Thr Ser Gln Met Glu Thr Phe His Ser Thr Ser Ser Glu Gly
                              265
                                                  270
          260
Asn Leu Ser Pro Ser Gln Val Ser Thr Thr Ser Glu His Pro Ser Gln
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Pro Ser Ser Pro Pro Asn Thr Thr Arg Gln
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                       295
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<212> DNA
<213> Respiratory syncytial virus
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<221> misc_feature
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<222> 9

 $\langle 223 \rangle$ n = A,T,C or G

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ccaaacaaac ccaataatga ttttcacttc gaaqtqttta actttgtacc ctgcagcata 540
tgcagcaaca atccaacctg ctgggctatc tgcaaaagaa taccaaacaa aaaaccagga 600
aagaaaacca ccaccaagcc tacaaaaaaa ccaaccttca agacaaccaa aaaagatcac 660
aaacctcaaa ccactaaacc aaaggaagta cccaccacca agcccacaga agagccaacc 720
atcaacacca ccaaaacaaa catcataact acactactca ccaacaacac cacaggaaat 780
ccaaaactca caagtcaaat ggaaaccttc cactcaacct cctccgaagg caatctaagc 840
cetteteaag tetecacaac ateegageac ceateacaac ceteatetee acceaacaca 900
acacgccagt agttatt
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Trp Asp Thr Leu Asn His Leu Leu Phe Ile Ser Ser Gly Leu Tyr Lys
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Leu Asn Leu Lys Ser Ile Ala Gln Ile Thr Leu Ser Ile Leu Ala Met
                           40
Ile Ile Ser Thr Ser Leu Ile Ile Thr Ala Ile Ile Phe Ile Ala Ser
                       55
Ala Asn His Lys Val Thr Leu Thr Thr Ala Ile Ile Gln Asp Ala Thr
                                      75
                   70
Ser Gln Ile Lys Asn Thr Thr Pro Thr Tyr Leu Thr Gln Asp Pro Gln
              85
Leu Gly Ile Ser Phe Ser Asn Leu Ser Glu Ile Thr Ser Gln Thr Thr
                               105
Thr Ile Leu Ala Ser Thr Thr Pro Gly Val Lys Ser Asn Leu Gln Pro
                           120
Thr Thr Val Lys Thr Lys Asn Thr Thr Thr Gln Thr Gln Pro Ser
                       135
                                           140
Lys Pro Thr Thr Lys Gln Arg Gln Asn Lys Pro Pro Asn Lys Pro Asn
                                      155
                  150
Asn Asp Phe His Phe Glu Val Phe Asn Phe Val Pro Cys Ser Ile Cys
              165
                                   170
Ser Asn Asn Pro Thr Cys Trp Ala Ile Cys Lys Arg Ile Pro Asn Lys
                               185
Lys Pro Gly Lys Lys Thr Thr Thr Lys Pro Thr Lys Lys Pro Thr Phe
                          200
Lys Thr Thr Lys Lys Asp His Lys Pro Gln Thr Thr Lys Pro Lys Glu
                      215
                                           220
Val Pro Thr Thr Lys Pro Thr Glu Glu Pro Thr Ile Asn Thr Thr Lys
                   230
                                      235
Thr Asn Ile Ile Thr Thr Leu Leu Thr Asn Asn Thr Thr Gly Asn Pro
              245
                                   250
Lys Leu Thr Ser Gln Met Glu Thr Phe His Ser Thr Ser Ser Glu Gly
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                               265
Asn Leu Ser Pro Ser Gln Val Ser Thr Thr Ser Glu His Pro Ser Gln
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Pro Ser Ser Pro Pro Asn Thr Thr Arg Gln
   290
                       295
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<210> 5 <211> 306

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<213> Artificial Sequence
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acaaaacaac gccaaaacaa accaccaaac aaacccaata atgattttca cttcgaagtg 120
tttaactttg taccctgcag catatgcagc aacaatccaa cctgctgggc tatctgcaaa 180
agaataccag ccaaaaaacc aggaaagaaa accaccacca agcctacaaa aaaaccaacc 240
ttcaagacaa ccaaaaaaga tcacaaacct caaaccacta aaccaaagga agtacccacc 300
accaag
<210> 6
<211> 102
<212> PRT
<213> Artificial Sequence
<220>
<223> RSV mutant fragment
<400> 6
Pro Thr Thr Val Lys Thr Lys Asn Thr Thr Thr Thr Gln Thr Gln Pro
Ser Lys Pro Thr Thr Lys Gln Arg Gln Asn Lys Pro Pro Asn Lys Pro
Asn Asn Asp Phe His Phe Glu Val Phe Asn Phe Val Pro Cys Ser Ile
                            40
Cys Ser Asn Asn Pro Thr Cyc Trp Ala Ile Cys Lys Arg Ile Pro Ala
                       55
Lys Lys Pro Gly Lys Lys Thr Thr Thr Lys Pro Thr Lys Lys Pro Thr
Phe Lys Thr Thr Lys Lys Asp His Lys Pro Gln Thr Thr Lys Pro Lys
Glu Val Pro Thr Thr Lys
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<210> 7
<211> 306
<212> DNA
<213> Artificial Sequence
<220>
<223> RSV mutant sequence
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acaaaacaac gccaaaacaa accaccaaac aaacccaata atgattttca cttcgaagtg 120
tttaactttg taccctgcag catatgcagc aacaatccaa cctgctgggc .tatctgcaaa 180
gcaataccaa acaaaaaacc aggaaagaaa accaccacca agcctacaaa aaaaccaacc 240
ttcaagacaa ccaaaaaaga tcacaaacct caaaccacta aaccaaagga agtacccacc 300
accaag
<210> 8
<211> 102
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                                    10
Ser Lys Pro Thr Thr Lys Gln Arg Gln Asn Lys Pro Pro Asn Lys Pro
                                25
Asn Asn Asp Phe His Phe Glu Val Phe Asn Phe Val Pro Cys Ser Ile
        35
                                               45
                            40
Cys Ser Asn Asn Pro Thr Cys Trp Ala Ile Cys Lys Ala Ile Pro Asn
                       55
                                            60
Lys Lys Pro Gly Lys Lys Thr Thr Lys Pro Thr Lys Lys Pro Thr
                 · 70
Phe Lys Thr Thr Lys Lys Asp His Lys Pro Gln Thr Thr Lys Pro Lys
                85
                                    90
Glu Val Pro Thr Thr Lys
            100
<210> 9
<211> 306
<212> DNA
<213> Artificial Sequence
<223> RSV mutant fragment
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tttaactttg taccotgoag catatgoago aacaatcoaa cotgotgggo tatotgoaaa 180
agaatagcaa acaaaaaacc aggaaagaaa accaccacca agcctacaaa aaaaccaacc 240
ttcaagacaa ccaaaaaaga tcacaaacct caaaccacta aaccaaagga agtacccacc 300
accaag
<210> 10
<211> 102
<212> PRT
<213> Artificial Sequence
<220>
<223> RSV mutant fragment
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                                    10
Ser Lys Pro Thr Thr Lys Gln Arg Gln Asn Lys Pro Pro Asn Lys Pro
                                25
Asn Asn Asp Phe His Phe Glu Val Phe Asn Phe Val Pro Cys Ser Ile
                                               45
                            40
Cys Ser Asn Asn Pro Thr Cys Trp Ala Ile Cys Lys Arg Ile Ala Asn
                       55
                                            60
Lys Lys Pro Gly Lys Lys Thr Thr Lys Pro Thr Lys Lys Pro Thr
                                        75
Phe Lys Thr Thr Lys Lys Asp His Lys Pro Gln Thr Thr Lys Pro Lys
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95 90 Glu Val Pro Thr Thr Lys 100 <210> 11 <211> 306 <212> DNA <213> Artificial Sequence <220> <223> RSV mutant fragment <400> 11 cccacaacag tcaagactaa aaacacaaca acaacccaaa cacaacccag caagcccact 60 acaaaacaac gccaaaacaa accaccaaac aaacccaata atgattttca cttcgaagtg 120 tttaactttg tacctgcag catatgcagc aacaatccaa cctgctgggc tgcctgcaaa 180 agaataccaa acaaaaaacc aggaaagaaa accaccacca agcctacaaa aaaaccaacc 240 ttcaagacaa ccaaaaaaga tcacaaacct caaaccacta aaccaaagga agtacccacc 300 accaag <210> 12 <211> 102 <212> PRT <213> Artificial Sequence <223> RSV mutant fragment <400> 12 Pro Thr Thr Val Lys Thr Lys Asn Thr Thr Thr Thr Gln Thr Gln Pro 10 Ser Lys Pro Thr Thr Lys Gln Arg Gln Asn Lys Pro Pro Asn Lys Pro 25 Asn Asn Asp Phe His Phe Glu Val Phe Asn Phe Val Pro Cys Ser Ile 40 Cys Ser Asn Asn Pro Thr Cys Trp Ala Ala Cys Lys Arg Ile Pro Asn Lys Lys Pro Gly Lys Lys Thr Thr Thr Lys Pro Thr Lys Lys Pro Thr 70 75 Phe Lys Thr Thr Lys Lys Asp His Lys Pro Gln Thr Thr Lys Pro Lys 85 Glu Val Pro Thr Thr Lys 100 <210> 13 <211> 306 <212> DNA <213> Artificial Sequence <220> <223> RSV mutant fragment <400> 13 cccacaacag tcaagactaa aaacacaaca acaacccaaa cacaacccag caagcccact 60 acaaaacaac gccaaaacaa accaccaaac aaacccaata atgattttca cttcgaagtg 120 tttaactttg taccctgcag catatgcagc aacaatccaa cctgctgggc tatcgccaaa 180

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agaataccaa acaaaaaacc aggaaagaaa accaccacca agcctacaaa aaaaccaacc 240
ttcaagacaa ccaaaaaaga tcacaaacct caaaccacta aaccaaagga agtacccacc 300
accaag
<210> 14
<211> 102
<212> PRT
<213> Artificial Sequence
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<223> RSV mutant fragment
<400> 14
Pro Thr Thr Val Lys Thr Lys Asn Thr Thr Thr Thr Gln Thr Gln Pro
                                    10
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                5
Ser Lys Pro Thr Thr Lys Gln Arg Gln Asn Lys Pro Pro Asn Lys Pro
                                25
Asn Asn Asp Phe His Phe Glu Val Phe Asn Phe Val Pro Cys Ser Ile
        35
                            40
Cys Ser Asn Asn Pro Thr Cys Trp Ala Ile Ala Lys Arg Ile Pro Asn
                                            60
                        55
Lys Lys Pro Gly Lys Lys Thr Thr Lys Pro Thr Lys Lys Pro Thr
                    70
                                        75
65
Phe Lys Thr Thr Lys Lys Asp His Lys Pro Gln Thr Thr Lys Pro Lys
                85
Glu Val Pro Thr Thr Lys
            100
<210> 15
<211> 306
<212> DNA
<213> Artificial Sequence
<220>
<223> RSV mutant fragment
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tttaactttg taccctgcag catatgcagc aacaatccaa cctgctgggc tatctgcgca 180
aquataccaa acaaaaaacc aggaaagaaa accaccacca agcctacaaa aaaaccaacc 240
ttcaagacaa ccaaaaaaga tcacaaacct caaaccacta aaccaaagga agtacccacc 300
                                                                  306
accaag
<210> 16
<211> 102
<212> PRT
<213> Artificial Sequence
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<400> 16
Pro Thr Thr Val Lys Thr Lys Asn Thr Thr Thr Gln Thr Gln Pro
                                   10
                                                       15
Ser Lys Pro Thr Thr Lys Gln Arg Gln Asn Lys Pro Pro Asn Lys Pro
            20
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Asn Asn Asp Phe His Phe Glu Val Phe Asn Phe Val Pro Cys Ser Ile
  35
                            40
                                               4.5
Cys Ser Asn Asn Pro Thr Cys Trp Ala Ile Cys Ala Arg Ile Pro Asn
                        55
Lys Lys Pro Gly Lys Lys Thr Thr Thr Lys Pro Thr Lys Lys Pro Thr
                                       75
Phe Lys Thr Thr Lys Lys Asp His Lys Pro Gln Thr Thr Lys Pro Lys
                85
                                    90
Glu Val Pro Thr Thr Lys
           100
<210> 17
<211> 306
<212> DNA
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acaaaacaac gccaaaacaa accaccaaac aaacccaata atgattttca cttcgaagtg 120
tttaactttg taccctgcag catatgcagc aacaatccaa cctgctgggc tatctgcaaa 180
agagcaccaa acaaaaaacc aggaaagaaa accaccacca agcctacaaa aaaaccaacc 240
ttcaagacaa ccaaaaaaga tcacaaacct caaaccacta aaccaaagga agtacccacc 300
<210> 18
<211> 102
<212> PRT
<213> Artificial Sequence
<220>
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<400> 18
Pro Thr Thr Val Lys Thr Lys Asn Thr Thr Thr Thr Gln Thr Gln Pro
                                    10
Ser Lys Pro Thr Thr Lys Gln Arg Gln Asn Lys Pro Pro Asn Lys Pro
                                25
Asn Asn Asp Phe His Phe Glu Val Phe Asn Phe Val Pro Cys Ser Ile
                            40
                                               45
Cys Ser Asn Asn Pro Thr Cys Trp Ala Ile Cys Lys Arg Ala Pro Asn
                       55
Lys Lys Pro Gly Lys Lys Thr Thr Lys Pro Thr Lys Lys Pro Thr
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Phe Lys Thr Thr Lys Lys Asp His Lys Pro Gln Thr Thr Lys Pro Lys
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<210> 19 <211> 306 <212> DNA <213> Artificial Sequence

Glu Val Pro Thr Thr Lys 100

85

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<220>
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tttaactttg taccctgcag catatgcagc aacaatccaa cctgctgggc tatctgcaaa 180
agaataccaa acgcaaaacc aggaaagaaa accaccacca agcctacaaa aaaaccaacc 240
ttcaagacaa ccaaaaaaga tcacaaacct caaaccacta aaccaaagga agtacccacc 300
accaag
<210> 20
<211> 102
<212> PRT
<213> Artificial Sequence
<220>
<223> RSV mutant fragment
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                                    10
Ser Lys Pro Thr Thr Lys Gln Arg Gln Asn Lys Pro Pro Asn Lys Pro
                                25
                                                    30
Asn Asn Asp Phe His Phe Glu Val Phe Asn Phe Val Pro Cys Ser Ile
                            40
                                                45
Cys Ser Asn Asn Pro Thr Cys Trp Ala Ile Cys Lys Arg Ile Pro Asn
                        55
Ala Lys Pro Gly Lys Lys Thr Thr Lys Pro Thr Lys Lys Pro Thr
                   70
                                        75
Phe Lys Thr Thr Lys Lys Asp His Lys Pro Gln Thr Thr Lys Pro Lys
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                                    90
Glu Val Pro Thr Thr Lys
            100
<210> 21
<211> 306
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acaaaacaac gccaaaacaa accaccaaac aaacccaata atgattttca cttcgaagtg 120
tttaactttg taccctgcag catatgcagc aacaatccaa cctgctgggc tatctgcaaa 180
agaataccaa acaaagcacc aggaaagaaa accaccacca agcctacaaa aaaaccaacc 240
ttcaagacaa ccaaaaaaga tcacaaacct caaaccacta aaccaaagga agtacccacc 300
accaag
<210> 22
<211> 102
<212> PRT
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<223> RSV mutant fragment
<400> 22
Pro Thr Thr Val Lys Thr Lys Asn Thr Thr Thr Gln Thr Gln Pro
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Ser Lys Pro Thr Thr Lys Gln Arg Gln Asn Lys Pro Pro Asn Lys Pro
            20
                                25
Asn Asn Asp Phe His Phe Glu Val Phe Asn Phe Val Pro Cys Ser Ile
                            40
Cys Ser Asn Asn Pro Thr Cys Trp Ala Ile Cys Lys Arg Ile Pro Asn
                        55
                                            60
Lys Ala Pro Gly Lys Lys Thr Thr Lys Pro Thr Lys Lys Pro Thr
65
                    70
                                        75
Phe Lys Thr Thr Lys Lys Asp His Lys Pro Gln Thr Thr Lys Pro Lys
                85
                                    90
Glu Val Pro Thr Thr Lys
            100
<210> 23
<211> 825
<212> DNA
<213> Artificial Sequence
<220>
<223> RSV mutant fragment
<221> CDS
<222> (1)...(825)
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Met Ser Asp Lys Ile Ile His Leu Thr Asp Asp Ser Phe Asp Thr Asp
                                     10
gta etc aaa geg gae ggg geg ate etc gte gat tte tgg gea gag tgg
                                                                   96
Val Leu Lys Ala Asp Gly Ala Ile Leu Val Asp Phe Trp Ala Glu Trp
             20
tgc ggt ccg tgc aaa atg atc gcc ccg att ctg gat gaa atc gct gac
Cys Gly Pro Cys Lys Met Ile Ala Pro Ile Leu Asp Glu Ile Ala Asp
gaa tat cag ggc aaa ctg acc gtt gca aaa ctg aac atc gat caa aac
                                                                   192
Glu Tyr Gln Gly Lys Leu Thr Val Ala Lys Leu Asn Ile Asp Gln Asn
                                                                  240
cet ggc act geg eeg aaa tat ggc ate egt ggt ate eeg act etg etg
Pro Gly Thr Ala Pro Lys Tyr Gly Ile Arg Gly Ile Pro Thr Leu Leu
ctg ttc aaa aac ggt gaa gtg gcg gca acc aaa gtg ggt gca ctg tct
                                                                  288
Leu Phe Lys Asn Gly Glu Val Ala Ala Thr Lys Val Gly Ala Leu Ser
aaa ggt cag ttg aaa gag ttc ctc gac gct aac ctg gcc ggt tct ggt
                                                                  336
Lys Gly Gln Leu Lys Glu Phe Leu Asp Ala Asn Leu Ala Gly Ser Gly
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105

110

100

			_							tct Ser			_			384
-				_		-		-	-	gct Ala			-	•	_	432
										gac Asp 155						480
										gaa Glu						528
-										caa Gln		-	-			576
			-							aaa Lys				-		624
		-	_				-		_	agc Ser		-	-			672
		-		-		-		-		cca Pro 235						720
					_					cca Pro			•			768
		•								cca Pro	_	-	-			816
acc Thr	aag Lys	tga *										٠				825

<210> 24

<211> 274

<212> PRT

<213> Artificial Sequence

<400> 24

 Met
 Ser
 Asp
 Lys
 Ile
 His
 Leu
 Thr
 Asp
 Asp
 Ser
 Phe
 Asp
 Thr
 Asp

 1
 5
 5
 10
 15
 15
 15

 Val
 Leu
 Leu
 Val
 Asp
 Phe
 Trp
 Ala
 Glu
 Trp

 20
 25
 25
 30
 30
 30

Cys Gly Pro Cys Lys Met Ile Ala Pro Ile Leu Asp Glu Ile Ala Asp

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Glu Tyr Gln Gly Lys Leu Thr Val Ala Lys Leu Asn Ile Asp Gln Asn 55 Pro Gly Thr Ala Pro Lys Tyr Gly Ile Arg Gly Ile Pro Thr Leu Leu 70 75 Leu Phe Lys Asn Gly Glu Val Ala Ala Thr Lys Val Gly Ala Leu Ser 90 85 Lys Gly Gln Leu Lys Glu Phe Leu Asp Ala Asn Leu Ala Gly Ser Gly 105 110 Ser Gly His Met His His His His His Ser Ser Gly Leu Val Pro 120 125 Arg Gly Ser Gly Met Lys Glu Thr Ala Ala Ala Lys Phe Glu Arg Gln 135 140 His Met Asp Ser Pro Asp Leu Gly Thr Asp Asp Asp Lys Thr Gly 150 155 Leu Leu Leu Asn His Gly Asp Ile Gly Ser Glu Phe Pro Thr Thr Val 170 165 Lys Thr Lys Asn Thr Thr Thr Thr Gln Thr Gln Pro Ser Lys Pro Thr 185 190 Thr Lys Gln Arg Gln Asn Lys Pro Pro Asn Lys Pro Asn Asn Asp Phe 200 His Phe Glu Val Phe Asn Phe Val Pro Cys Ser Ile Cys Ser Asn Asn 215 220 Pro Thr Cys Trp Ala Ile Cys Lys Arg Ile Pro Asn Lys Lys Pro Gly 230 235 Lys Lys Thr Thr Thr Lys Pro Thr Lys Lys Pro Thr Phe Lys Thr Thr 245 250 Lys Lys Asp Leu Lys Pro Gln Thr Thr Lys Pro Lys Glu Val Pro Thr 265 Thr Lys

<210> 25

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<212> DNA

<213> Artificial Sequence

<220>

<223> RSV mutant fragment

<221> CDS

<222> (1) ... (825)

<400> 25

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gta ctc aaa gcg gac ggg gcg atc ctc gtc gat ttc tgg gca gag tgg 96 Val Leu Lys Ala Asp Gly Ala Ile Leu Val Asp Phe Trp Ala Glu Trp

tgc ggt ccg tgc aaa atg atc gcc ccg att ctg gat gaa atc gct gac 144 Cys Gly Pro Cys Lys Met Ile Ala Pro Ile Leu Asp Glu Ile Ala Asp 35 40

gaa tat cag ggc aaa ctg acc gtt gca aaa ctg aac atc gat caa aac 192 Glu Tyr Gln Gly Lys Leu Thr Val Ala Lys Leu Asn Ile Asp Gln Asn

	50					55					60					
cct	ggc	act	gcg	ccg	aaa	tat	ggc	atc	cgt	ggt	atc	ccg	act	ctg	ctg	240
Pro 65	GTÀ	Thr	Ala	Pro	Lys 70	Tyr	Gly	IIe	Arg	75	11e	PIO	Thr	Leu	80	
ctg Leu	ttc Phe	aaa Lys	aac Asn	ggt Gly 85	gaa Glu	gtg Val	gcg Ala	gca Ala	acc Thr 90	aaa Lys	gtg Val	ggt Gly	gca Ala	ctg Leu 95	tct Ser	288
							ctc Leu									336
			-				cat His 120						_			384
							acc Thr									432
cac His 145	atg Met	gac Asp	agc Ser	cca Pro	gat Asp 150	ctg Leu	ggt Gly	acc Thr	gat Asp	gac Asp 155	gac Asp	gac Asp	aag Lys	acc Thr	ggg Gly 160	480
							atc Ile									528
							acc Thr									576
							cca Pro 200									624
							gta Val									672
							aaa Lys									720
							aca Thr									768
							acc Thr									816
acc Thr	aag Lys	tga *														825

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<210> 26
<211> 274
<212> PRT
<213> Artificial Sequence
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Val Leu Lys Ala Asp Gly Ala Ile Leu Val Asp Phe Trp Ala Glu Trp
                               25
Cys Gly Pro Cys Lys Met Ile Ala Pro Ile Leu Asp Glu Ile Ala Asp
                           40
Glu Tyr Gln Gly Lys Leu Thr Val Ala Lys Leu Asn Ile Asp Gln Asn
   50
                       55
                                           60
Pro Gly Thr Ala Pro Lys Tyr Gly Ile Arg Gly Ile Pro Thr Leu Leu
                   70
                                       75
Leu Phe Lys Asn Gly Glu Val Ala Ala Thr Lys Val Gly Ala Leu Ser
                                   90
Lys Gly Gln Leu Lys Glu Phe Leu Asp Ala Asn Leu Ala Gly Ser Gly
                               105
Ser Gly His Met His His His His His His Ser Ser Gly Leu Val Pro
                           120
                                               125
Arg Gly Ser Gly Met Lys Glu Thr Ala Ala Ala Lys Phe Glu Arg Gln
                      135
                                           140
His Met Asp Ser Pro Asp Leu Gly Thr Asp Asp Asp Lys Thr Gly
                   150
                                       155
Leu Leu Leu Asn His Gly Asp Ile Gly Ser Glu Phe Pro Thr Thr Val
                                   170
                                                       175
              165
Lys Thr Lys Asn Thr Thr Thr Gln Thr Gln Pro Ser Lys Pro Thr
                               185
                                                   190
Thr Lys Gln Arg Gln Asn Lys Pro Pro Asn Lys Pro Asn Asn Asp Phe
                           200
                                               205
His Phe Glu Val Phe Asn Phe Val Pro Cys Ser Ile Cys Ser Asn Asn
                       215
                                           220
Pro Thr Cys Trp Ala Ile Cys Lys Arg Ile Pro Ala Lys Lys Pro Gly
                                      235
                  230
Lys Lys Thr Thr Thr Lys Pro Thr Lys Lys Pro Thr Phe Lys Thr Thr
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                                  250
Lys Lys Asp Leu Lys Pro Gln Thr Thr Lys Pro Lys Glu Val Pro Thr
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Thr Lys
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<210> 27
<211> 852
<212> DNA
<213> Artificial Sequence
<220>
<223> RSV mutant fragment
<221> CDS
<222> (1)...(852)
<400> 27
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-		-	-	_		_	_	-	-				-	G] y		96
		_	-			-			-		-	-		atg Met		144
														ctg Leu		192
_	_		_			_							_	aaa Lys		240
		-			-		-	_	-					gaa Glu 95		288
														gag Glu		336
	_	_		-	_							-		cat His		384
						_			-				_	aaa Lys	-	432
	-	•	_			-	_	-		_	-	_		gat Asp	-	480
		-	_	-	-	_								ggc Gly 1 75	•	528
			-					-						aca Thr		576
														aac Asn		624
							_				-			aac Asn		672
														atc Ile		720

275

768 Lys Arg Ile Pro Asn Lys Lys Pro Gly Lys Lys Thr Thr Thr Lys Pro aca aaa aaa cca acc ttc aag aca acc aaa aaa gat ctc aaa cct caa Thr Lys Lys Pro Thr Phe Lys Thr Thr Lys Lys Asp Leu Lys Pro Gln 265 852 acc act aaa cca aag gaa gta ccc acc acc aag tga Thr Thr Lys Pro Lys Glu Val Pro Thr Thr Lys * 275 <210> 28 <211> 283 <212> PRT <213> Artificial Sequence <400> 28 Met Phe Leu Leu Ala Val Phe Tyr Gly Gly Ser Asp Lys Ile Ile His 10 Leu Thr Asp Asp Ser Phe Asp Thr Asp Val Leu Lys Ala Asp Gly Ala 25 Ile Leu Val Asp Phe Trp Ala Glu Trp Cys Gly Pro Cys Lys Met Ile 40 Ala Pro Ile Leu Asp Glu Ile Ala Asp Glu Tyr Gln Gly Lys Leu Thr Val Ala Lys Leu Asn Ile Asp Gln Asn Pro Gly Thr Ala Pro Lys Tyr 70 Gly Ile Arg Gly Ile Pro Thr Leu Leu Leu Phe Lys Asn Gly Glu Val 90 Ala Ala Thr Lys Val Gly Ala Leu Ser Lys Gly Gln Leu Lys Glu Phe 105 Leu Asp Ala Asn Leu Ala Gly Ser Gly Ser Gly His Met His His His 120 His His Ser Ser Gly Leu Val Pro Arg Gly Ser Gly Met Lys Glu 135 Thr Ala Ala Ala Lys Phe Glu Arg Gln His Met Asp Ser Pro Asp Leu 150 155 Gly Thr Asp Asp Asp Asp Lys Thr Gly Leu Leu Asn His Gly Asp 170 175 Ile Gly Ser Glu Phe Pro Thr Thr Val Lys Thr Lys Asn Thr Thr 185 190 Thr Gln Thr Gln Pro Ser Lys Pro Thr Thr Lys Gln Arg Gln Asn Lys 200 Pro Pro Asn Lys Pro Asn Asn Asp Phe His Phe Glu Val Phe Asn Phe 215 220 Val Pro Cys Ser Ile Cys Ser Asn Asn Pro Thr Cys Trp Ala Ile Cys 235 230 Lys Arg Ile Pro Asn Lys Lys Pro Gly Lys Lys Thr Thr Thr Lys Pro 250 255 Thr Lys Lys Pro Thr Phe Lys Thr Thr Lys Lys Asp Leu Lys Pro Gln 265 Thr Thr Lys Pro Lys Glu Val Pro Thr Thr Lys

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-	-		-			-	caa Gln					_		240
		_			-		ctg Leu	-	_				-	 288
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gta ccc tgc ago Val Pro Cys Ser 225	_	_				-		-		-	720
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Glu	Tyr 50	35 Gln	Gly	Lys	Leu	Thr 55	40 Val	Ala	Lys	Leu	Asn 60	45 Ile	Asp	Gln	Asn	
Pro 65		Thr	Ala	Pro	Lys 70		Gly	Ile	Arg	Gly 75	Ile	Pro	Thr	Leu	Leu 80	
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Lys Gly Gln Leu Lys Glu Phe Leu Asp Ala Asn Leu Ala Gly Ser Gly 105 110 100 Ser Gly His Met His His His His His Ser Ser Gly Leu Val Pro 125 120 Arg Gly Ser Gly Met Lys Glu Thr Ala Ala Ala Lys Phe Glu Arg Gln 135 140 His Met Asp Ser Pro Asp Leu Gly Thr Asp Asp Asp Lys Thr Gly 155 150 Leu Leu Leu Asn His Gly Asp Ile Gly Ser Glu Phe Pro Thr Thr Val 170 165 Lys Thr Lys Asn Thr Thr Thr Gln Thr Gln Pro Ser Lys Pro Thr 185 190 Thr Lys Gln Arg Gln Asn Lys Pro Pro Asn Lys Pro Asn Asn Asp Phe 200 205 His Phe Glu Val Phe Asn Phe Val Pro Cys Ser Ile Cys Ser Asn Asn 210 215 220 Pro Thr Cys Trp Ala Ile Cys Lys Arg Ile Pro Asn Lys Lys Pro Gly 235 230 Lys Lys Thr Thr Thr Lys Pro Thr Lys Lys Pro Thr Phe Lys Thr Thr 250 245 Lys Lys Asp Leu Lys Pro Gln Thr Thr Lys Pro Lys Glu Val Pro Thr 265 Thr Lys Gly Gly Tyr Phe Val Ala Leu Leu Phe 280 <210> 33 <211> 852 <212> DNA <213> Artificial Sequence <220> <223> RSV mutant fragment <221> CDS <222> (1)...(852) atg tee gae aaa ate ate cae etg act gae gae agt tit gae aeg gat 48 Met Ser Asp Lys Ile Ile His Leu Thr Asp Asp Ser Phe Asp Thr Asp 5 gta ctc aaa gcg gac ggg gcg atc ctc gtc gat ttc tgg gca gag tgg Val Leu Lys Ala Asp Gly Ala Ile Leu Val Asp Phe Trp Ala Glu Trp tgc ggt eeg tgc aaa atg atc gec eeg att etg gat gaa atc get gac 144 Cys Gly Pro Cys Lys Met Ile Ala Pro Ile Leu Asp Glu Ile Ala Asp 40 192 gaa tat cag ggc aaa ctg acc gtt gca aaa ctg aac atc gat caa aac Glu Tyr Gln Gly Lys Leu Thr Val Ala Lys Leu Asn Ile Asp Gln Asn 50 cct ggc act gcg ccg aaa tat ggc atc cgt ggt atc ccg act ctg ctg Pro Gly Thr Ala Pro Lys Tyr Gly Ile Arg Gly Ile Pro Thr Leu Leu

75

70

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 tttaactttg taccctgcag catatgcagc gccaatccaa cctgctgggc tatctgcaaa 180
 agaataccaa acaaaaaacc aggaaagaaa accaccacca agcctacaaa aaaaccaacc 240
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                             40
Cys Ser Ala Asn Pro Thr Cys Trp Ala Ile Cys Lys Arg Ile Pro Asn
                         55
Lys Lys Pro Gly Lys Lys Thr Thr Lys Pro Thr Lys Lys Pro Thr
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Phe Lys Thr Thr Lys Lys Asp His Lys Pro Gln Thr Thr Lys Pro Lys
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Glu Val Pro Thr Thr Lys
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Asp Leu Tyr Asp Asp Asp Lys

Application No PCT/CA2004/001007

A. CLASSIFICATION OF SUBJECT MATTER
IPC 7 A61K39/155 C07K14/135 A61K39/39 C12N15/45

C07K16/10

C12N15/62

A61K39/295

Relevant to claim No.

According to International Patent Classification (IPC) or to both national classification and IPC

B. FIELDS SEARCHED

Category °

Minimum documentation searched (classification system followed by classification symbols)

IPC 7 A61K C07K

Documentation searched other than minimum documentation to the extent that such documents are included in the fields searched

Electronic data base consulted during the international search (name of data base and, where practical, search terms used)

Citation of document, with indication, where appropriate, of the relevant passages

EPO-Internal, BIOSIS, EMBASE, WPI Data

C. DOCUMENTS CONSIDERED TO BE RELEVANT

Х	WO 99/14334 A (TEBBEY PAUL W ; / CYANAMID CO (US); HANCOCK GERAL 25 March 1999 (1999-03-25) the whole document		1-45
X	HUANG Y ET AL: "A single amino substitution in a recombinant 6 vaccine drastically curtails profimmunity against respiratory synvirus (RSV)" VACCINE, BUTTERWORTH SCIENTIFIC. GUILDFORD, GB, vol. 21, no. 19-20, 2 June 2003 (2003-06-02), pages XP004424161 ISSN: 0264-410X the whole document	protein otective ocytial	1-45
X Furth	er documents are listed in the continuation of box C.	Patent family members are listed in	n annex.
'A' docume conside 'E' earlier diffiling de 'L' docume which i citation 'O' docume other n' 'P' docume	nt which may throw doubts on priority claim(s) or s cited to establish the publication date of another or other special reason (as specified) ont referring to an oral disclosure, use, exhibition or	 'T' later document published after the interest or priority date and not in conflict with cited to understand the principle or the invention 'X' document of particular relevance; the cleannot be considered novel or cannot involve an inventive step when the document of particular relevance; the cleannot be considered to involve an inventive step when the document is combined with one or moments, such combination being obvious in the art. '8' document member of the same patent for the considered to the same patent for the same patent f	the application but ory underlying the latmed invention be considered to current is taken alone aimed invention rentive step when the re other such docu-s to a person skilled
Date of the a	ctual completion of the international search	Date of mailing of the international sear	ch report
19	November 2004	08/12/2004	
Name and m	hailing address of the ISA European Palent Office, P.B. 5618 Patentlaan 2 NL - 2260 HV Rljswijk Tel. (+31-70) 340-2040, Tx. 31 651 epo nl, Fax: (+31-70) 340-3016	Authorized officer Sommerfeld, T	

Intermal Application No
PCT/CA2004/001007

		PCT/CA2004/001007
<u> </u>	ation) DOCUMENTS CONSIDERED TO BE RELEVANT	
ategory °	Cliation of document, with Indication, where appropriate, of the relevant passages	Relevant to claim No.
Y	HUANG Y ET AL: "Enhanced immune protection by a liposome-encapsulated recombinant respiratory syncytial virus (RSV) vaccine using immunogenic lipids from Deinococcus radiodurans" VACCINE, BUTTERWORTH SCIENTIFIC. GUILDFORD, GB, vol. 20, no. 11-12, 22 February 2002 (2002-02-22), pages 1586-1592, XP004340380 ISSN: 0264-410X the whole document	1-45
Y	SPARER T E ET AL: "Eliminating a region of respiratory syncytial virus attachment protein allows induction of protective immunity without vaccine-enhanced lung eosinophilia" JOURNAL OF EXPERIMENTAL MEDICINE, TOKYO, JP, vol. 187, no. 11, 1 June 1998 (1998-06-01), pages 1921-1926, XP002091345 ISSN: 0022-1007 the whole document	1-45
	VARGA S M ET AL: "The attachment (G) glycoprotein of respiratory syncytial virus contains a single immunodominant epitope that elicits both Th1 and Th2 CD4+ T cell responses." JOURNAL OF IMMUNOLOGY (BALTIMORE, MD.: 1950) 1 DEC 2000, vol. 165, no. 11, 1 December 2000 (2000-12-01), pages 6487-6495, XP002306390 ISSN: 0022-1767 the whole document	1-45
•	SRIKIATKHACHORN ANON ET AL: "Induction of Th-1 and Th-2 responses by respiratory syncytial virus attachment glycoprotein is epitope and major histocompatibility complex independent" JOURNAL OF VIROLOGY, vol. 73, no. 8, August 1999 (1999-08), pages 6590-6597, XP002306391 ISSN: 0022-538X the whole document	1-45

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		PCT/CA2004/001007				
(Continuation) DOCUMENTS CONSIDERED TO BE RELEVANT						
Calegory "	Citation of document, with indication, where appropriate, of the relevant passages		Relevant to daim No.			
Υ	TEBBEY PW ET AL: "Atypical pulmonary eosinophilla is mediated by a specific amino acid sequence of the atachment (G) protein of respiratory syncytial virus" JOURNAL OF EXPERIMENTAL MEDICINE, TOKYO, JP, vol. 188, no. 10, 16 November 1998 (1998-11-16), pages 1967-1972, XP002091346 ISSN: 0022-1007 the whole document					
Y	WO 02/058725 A (PF MEDICAMENT; POWER ULTAN (FR); N GUYEN THIEN NGOC (FR)) 1 August 2002 (2002-08-01) the whole document		1-45			
A	RYAN E J ET AL: "Immunomodulators and delivery systems for vaccination by mucosal routes" TRENDS IN BIOTECHNOLOGY, ELSEVIER, AMSTERDAM,, GB, vol. 19, no. 8, 1 August 2001 (2001-08-01), pages 293-304, XP004254296 ISSN: 0167-7799 the whole document					
A	WO 01/60402 A (INTELLIVAX INTERNAT INC; WHITE GREGORY LEE (CA); PLANTE MARTIN (CA);) 23 August 2001 (2001-08-23) the whole document					

PCT/CA2004/001007

Box II Observations where certain claims were found unsearchable (Continuation of Item 2 of first sheet)
This International Search Report has not been established in respect of certain claims under Article 17(2)(a) for the following reasons:
1. X Claims Nos.: because they relate to subject matter not required to be searched by this Authority, namely:
Although claims 1-28 and 30 are directed to a method of treatment of the human/animal body, the search has been carried out and based on the alleged effects of the compound/composition.
Claims Nos.: because they relate to parts of the International Application that do not comply with the prescribed requirements to such an extent that no meaningful International Search can be carried out, specifically:
3. Claims Nos.: because they are dependent claims and are not drafted in accordance with the second and third sentences of Rule 6.4(a).
Box III Observations where unity of invention is lacking (Continuation of item 3 of first sheet)
This International Searching Authority found multiple inventions in this international application, as follows:
As all required additional search fees were timely paid by the applicant, this International Search Report covers all searchable claims.
As all searchable claims could be searched without effort justifying an additional fee, this Authority did not invite payment of any additional fee.
3. As only some of the required additional search fees were timely paid by the applicant, this International Search Report covers only those claims for which fees were paid, specifically claims Nos.:
No required additional search fees were timely paid by the applicant. Consequently, this International Search Report is restricted to the invention first mentioned in the claims; it is covered by claims Nos.:
Remark on Protest The additional search fees were accompanied by the applicant's protest.
No protest accompanied the payment of additional search fees.

Information on patent family members

Interreshal Application No PCT/CA2004/001007

Patent document cited in search report		Publication date		Patent family member(s)	Publication date
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			WO	0160402 A2	23-08-2001
			ÜS	2004156867 A1	12-08-2004
			ŪŠ	2001053368 A1	20-12-2001

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